

EFFECTS OF APPLICATION METHODS AND RATES OF MALEIC HYDRAZIDE ON THE COMPOSITION OF BURLEY TOBACCO



Mingwu Cui², H.R. Burton^{1,2}, L.P. Bush², T.G. Sutton³, and S.J. Crafts-Brandner³

A two-year study was completed to determine the effects of application methods and rates of maleic hydrazide (MH) on the composition of burley tobacco. MH applications were made at 0.5, 1.0, 1.5, 2.0 times the recommended, labeled rate (170mg plant⁻¹) using three application methods (single, split, and reduced-volume application). With an increased rate of MH application, calcium, magnesium, phosphorus, alkaloids, and nitrate levels were decreased, whereas potassium levels were increased. α -4,8,13-duvatriene-1,3-diol increased when MH applications exceeded the

recommended rate, whereas β -4,8,13-duvatriene-1,3-diol and solanesol level were not significantly changed. The different application methods affected MH residues, but they had no significant effects on leaf composition. No significant differences were measured in the accumulation of dry matter in leaves from the entire plant.

Additional key words: *Nicotiana tabacum* L., maleic hydrazide, leaf composition, cations, anions, alkaloid, solanesol, diterpenediols.

INTRODUCTION

Topping (decapitation) and sucker control (inhibition or removal of axillary buds) are two key cultural practices for producing quality tobacco (*Nicotiana tabacum* L.). Maleic hydrazide (1,2-dihydro-3,6-pyridazinedione) (MH) is a systemic plant growth regulator that has been the most effective and most extensively used sucker control agent for tobacco in the United States. The influence of MH on the agronomic, physical, and chemical properties of cured tobacco has been investigated extensively (7,10,11,38,44,45,48). In general, MH results in higher yields, equilibrium moisture, sugar and K⁺, but decreased nicotine and nitrate when compared to hand-suckered controls. However, studies have not examined changes of leaf components as affected by different application methods and rates of MH. There is an absence of information on the effects of MH application on solanesol and the leaf surface diterpenes, α - and β -4,8,13-duvatriene-1,3-diols (DVT), of burley tobacco. The objective of this study was to determine changes in MH residues and major chemical components in leaves of burley tobacco after different MH application rates and methods were used.

MATERIALS AND METHODS

Burley tobacco (cv. KY14) was grown using recommended practices at the Kentucky Agricultural Experiment Station Farm (Spindletop) at Lexington, Ky., in 1989 and 1990. The fertilization and production practices for this study have been described (10,11). The following treatments were replicated four times in a randomized complete block design:

A. Control.

1. Hand-suckered control.

B. Single applications of MH with recommended volume of water (392 L ha⁻¹ or 20 mL plant⁻¹).

2. 85 mg MH plant⁻¹ (0.5x, x = recommended rate of MH).

3. 170 mg MH plant⁻¹ (1.0 x).

4. 255 mg MH plant⁻¹ (1.5 x).

5. 340 mg MH plant⁻¹ (2.0 x).

C. Split-application of MH with recommended volume of water (392 L ha⁻¹ or 20 mL plant⁻¹).

6. 85 mg MH plant⁻¹ + 85 mg plant⁻¹ (0.5 x + 0.5 x).

7. 170 mg MH plant⁻¹ + 170 mg plant⁻¹ (1.0 x + 1.0 x).

8. 255 mg MH plant⁻¹ + 255 mg plant⁻¹ (1.5 x + 1.5 x).

9. 340 mg MH plant⁻¹ + 340 mg plant⁻¹ (2.0 x + 2.0 x).

D. Single application of MH with one-half the recommended volume of water (196 L ha⁻¹ or 10 mL plant⁻¹).

¹To whom correspondence should be addressed.

²Department of Agronomy, University of Kentucky, Lexington, KY 40546.

³USDA, ARS, University of Kentucky, Lexington, KY, 40546.

10. 170 mg MH plant⁻¹ (1.0 x).
11. 340 mg MH plant⁻¹ (2.0 x).
12. 170 mg MH plant⁻¹ + 170 mg plant⁻¹ (1.0 x + 1.0 x).

Green samples from plants that received the recommended rate of MH (treatment 3) were taken at four-day intervals to determine changes in MH residues from time of application to harvest. Each green sample consisted of two leaves from the top stalk position, with one leaf from each of two plants. Midribs of the leaves were removed from the green samples. Lamina tissue was freeze-dried and then ground to pass through a 850- μ m screen before residue analysis. Tobacco plants were stalk-cut on the 25th day after topping and air-cured in a conventional barn. Cured leaves were removed from stalks and separated into three leaf positions (top, middle, and bottom) for determination of leaf yields. Ten leaves were randomly pulled from each position for determination of dry matter and chemical analyses. Lamina and midribs were separated, dried at 65°C, weighed, ground to pass through an 850- μ m screen, and stored at ambient temperature in the dark before chemical analyses. Stalks were dried in an oven at 65°C for determination of dry matter.

Maleic hydrazide was determined by the AOAC procedure described by Lane (28). Ground tobacco was washed with 9:1 nitric acid:perchloric acid for analyses of potassium, calcium, and magnesium by atomic absorption spectroscopy (24). Soluble phosphate, sulfate, and chloride were determined by ion chromatography with a Dionex model 4000i equipped with AS4A separation column, AG4A guard column and a conductivity detector with background suppression. Total nitrogen was determined, following micro Kjeldahl block digestion, by the phenol-hypochlorite procedure using a Technicon Autoanalyzer (4). Total phosphorus was determined simultaneously on the digested sample using the method of Fiske & Subbarow (18). Nitrate-N was measured with an Automated Technicon Analyzer-System II by Griess reaction after Cd-Cu reduction of NO₃- to NO₂- (46). Individual alkaloids, nicotine, nornicotine, anabasine, anatabine, and myosmine were determined by the method described by Severson et al. (41) and modified by Madsen et al. (31). The values for total alkaloids represented a sum of the individual alkaloids. Solanesol and α - and β -4,8,13-duvatriene-1,3-diols were determined by a GC

procedure described by Burton et al. (5) and Severson et al. (40). Nitrite-N was determined by using the Griess procedure reported by Crutchfield & Burton (12).

Data analyses were performed as outlined by the SAS Institute (36). Year and replications were combined as blocks, positions were treated as split-plots and polynomial regression was applied to MH residue vs application rates within each application method. Single degree of freedom contrasts were used to test the mean difference of residues from different application methods with same level of active ingredient. LSD was used to compare means.

RESULTS AND DISCUSSION

Agronomic and Physical Characteristics

Across all application rates and methods, MH-treated plants did not have appreciable sucker growth during the first 21 days after topping. A few small suckers developed after 21 days in the treatments with lowest MH rates (85 mg MH plant⁻¹). The values for leaf dry weight and the moisture content tended to be slightly higher and stalk dry weight tended to be lower for MH-treated tobacco compared to hand-suckered tobacco; however, none of these differences was significant (data not shown). Some studies have reported increased yields of MH-treated tobacco over hand-suckered tobacco (38,44,45), but others (1,29,33,35,42) reported that yields either decreased or were not affected.

MH Residues

Following the recommended application rate (170 mg MH plant⁻¹), MH residues of the top leaves decreased with time after application (Figure 1), and they best fit the power model (R = 0.99**):

$$\text{MH} = 502 \times \text{Day}^{-0.84} \quad (1)$$

where Day = days after MH application, and MH is in $\mu\text{g g}^{-1}$.

Residue levels decreased greatly in the first eight days and were in the 30-70 $\mu\text{g g}^{-1}$ range by 12 days after treatment. There was no further significant decrease in MH levels after 12 days. Previous studies showed that lengthening the time between application of MH and harvest usually resulted in lower MH residues (9,14,22,38). Because stalk-cutting of burley tobacco is normally 3-4 weeks after topping, our data indicated that a further delay of harvest to reduce MH residue on burley

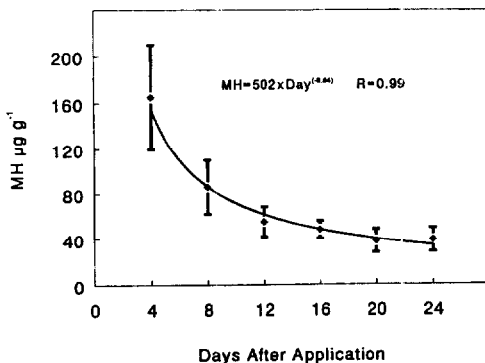
tobacco would not be effective when the recommended application rate and method were followed.

MH residues increase with increased amount of application (14,38). Our results confirm these observations, but to compare the different application methods, only treatments of the same application rate are summarized in Table 1. Treatments with split-application of MH and with reduced volume of water (high concentration of MH) had higher residue levels than the recommended single application. The effect of split-application has been reported (13) and the high MH residue levels can be explained by the second application being seven days closer to harvest than the single application. Because the potassium salt of MH is very water soluble, it can be assumed that by harvest most of the residue on the leaf surface had been removed by rain and heavy dews. Therefore, the increased MH residue from the single reduced-water application may be a result of more absorption because of its higher concentration in the solution. Furthermore, MH residues in green samples and in cured midribs (data not shown) have the same patterns of residue changes from the different application methods and application rates, which supports the hypothesis of greater MH absorption by the plant from the reduced water-volume application.

Chemical Composition

Whole-plant averages of selected chemical constituents in cured lamina for all treatments

Figure 1. Changes in residues of maleic hydrazide (MH) in green leaf samples of the top stalk position of burley tobacco with increased time after application of 170 mg plant⁻¹ MH. The analysis was conducted on lamina tissue. Each value is the mean of eight replications from a two-year study and the vertical bars represent the standard errors of the means.



are shown in Table 2. The type of MH treatment had a significant effect on Ca²⁺, Mg²⁺, K⁺, P, NO₃-N, and total alkaloid composition, while no treatment effects were measured for NO₂-N and total N composition. Compared to the hand-suckered control, MH-treated tobacco had increased K⁺ (17-34% higher), and decreased Ca²⁺ (9-16% lower), Mg²⁺ (4-16% lower), phosphorus (3-11% lower), total alkaloids (8-26% lower), and NO₃-N (8-28% lower). When the application rate of

Table 1. Maleic hydrazide residues at three stalk positions on air-cured burley tobacco after different application methods at equal application rates.

Method	Treatment		Stalk position		
	Water volume	MH rate	Top	Middle	Bottom
	--- L ha ⁻¹ ---	--- kg ha ⁻¹ ---	----- µg g ⁻¹ -----		
Single	392	3.36	23.4	24.3	16.9
Single	392	6.72	127.7	58.2	38.4
		Mean	75.6	41.2	27.7
Split	392	1.68+1.68	52.4	34.8	24.7
Split	392	3.76+3.76	185.5	78.7	42.2
		Mean	119.0	56.7	33.5
Single	196	3.36	55.6	27.3	25.5
Single	196	6.72	205.1	104.9	43.1
		Mean	132.4	66.1	34.3
		LSD _{0.05} ^a	40.5	ns	ns

^aLSDs apply to means only.

Table 2. Effects of application methods and rates of maleic hydrazide on selected chemical constituents in air-cured lamina of burley tobacco (whole plant averages).

Method	Treatment		Chemical constituent								
	Water volume	MH rate	Ca ²⁺	K ⁺	Mg ²⁺	P	NO ₃ -N	Total N	Total alkaloids	NO ₂ -N	
		- L ha ⁻¹ -	----- mg g ⁻¹ -----								µg g ⁻¹
Control	---	---	43.1	24.2	7.6	2.9	5.2	39.2	38.1	2.7	
Single	392	1.68	39.3	28.6	7.3	2.8	4.8	40.2	32.5	2.7	
Single	392	3.36	38.3	29.7	6.8	2.7	4.5	40.3	35.1	2.4	
Single	392	5.04	37.6	29.0	6.7	2.7	4.2	41.4	31.7	2.5	
Single	392	6.72	37.2	31.2	6.6	2.7	4.4	41.5	30.0	2.5	
Split	392	1.68+1.68	37.0	28.3	6.9	2.7	4.4	40.7	32.3	2.7	
Split	392	3.36+3.36	36.4	28.8	6.4	2.7	4.0	40.2	29.0	2.7	
Split	392	5.04+5.04	36.5	29.5	6.5	2.7	3.5	41.6	28.8	2.7	
Split	392	6.72+6.72	36.1	30.7	6.5	2.6	3.9	41.0	28.5	2.5	
Single	196	3.36	38.4	29.7	6.7	2.7	4.0	39.8	33.6	2.6	
Single	196	6.72	37.7	32.4	6.6	2.6	3.8	40.2	28.7	2.7	
Split	196	3.36+3.36	36.9	30.6	6.6	2.7	3.7	40.1	31.2	2.7	
LSD _{0.05}			1.5	2.4	0.3	0.1	0.5	ns	3.1	ns	

MH was increased within the same application method, changes of chemical constituents were small or non-significant and inconsistent. Although significant differences for chemical composition were found between the 0.5x rate and higher rates, the largest change was usually between the hand-suckered and 0.5x rate of MH application. F-tests of the single degree freedom contrasts of the changes in chemical composition among different application methods with equivalent rates of the active ingredient were not significant.

Means for each component at each stalk position of MH-treated tobacco and the hand-

suckered control are presented in **Table 3**. The changes in composition of lamina from different stalk positions of MH-treated tobacco and the hand-suckered control were very similar. In the cured midrib, each of the constituents was not significantly affected by MH application on a whole plant basis or by leaves within stalk position (data not shown).

The major cations (Ca²⁺, Mg²⁺, and K⁺) and anions (P and NO₃-N) in leaf lamina from each stalk position are presented in **Table 4**. Like the effect of MH application on MH residue levels, most of the significant differences in ion content were measured from the top stalk

Table 3. Effects of stalk positions on chemical constituents in air-cured lamina of maleic hydrazide-treated and hand-suckered burley tobacco.

Stalk positions	Chemical constituent									
	Ca ²⁺	K ⁺	Mg ²⁺	P	NO ₃ -N	Total N	Total alkaloids	NO ₂ -N		
		----- mg g ⁻¹ -----								µg g ⁻¹
MH-Treated										
Top	28.6	29.8	5.6	2.6	1.7	44.5	34.9	2.7		
Middle	38.0	29.7	6.7	2.8	4.2	42.8	36.1	2.6		
Bottom	47.5	28.6	7.9	2.7	6.7	34.3	23.3	2.6		
LSD _{0.05}	1.0	ns	0.2	0.1	1.1	2.2	2.2	ns		
Hand-Suckered										
Top	35.2	23.7	6.9	3.1	2.3	44.2	43.8	2.7		
Middle	42.0	25.2	7.3	3.0	5.0	41.1	41.5	2.6		
Bottom	49.8	23.8	8.5	2.7	8.3	33.2	28.3	2.6		
LSD _{0.05}	1.2	ns	0.4	0.2	1.9	2.8	2.5	ns		

Table 4. Effects of application methods and rates of maleic hydrazide on major cations and anions in air-cured lamina of burley tobacco.

Treatment			Chemical Constituent														
			Ca ²⁺			Mg ²⁺			K ⁺			P			NO ₃ -N		
Method	Water volume	MH rate	Top	Mid.	Bot.	Top	Mid.	Bot.	Top	Mid.	Bot.	Top	Mid.	Bot.	Top	Mid.	Bot.
	L ha ⁻¹	kg ha ⁻¹	mg g ⁻¹														
Control	----	----	5.2	42.0	49.2	6.9	7.3	8.5	23.6	25.2	23.8	3.1	3.1	2.7	2.4	5.0	8.1
Single	392	1.68	31.9	39.8	49.2	6.4	7.1	8.3	28.9	27.6	29.1	2.9	3.0	2.5	2.1	4.9	7.9
Single	392	3.36	28.7	38.4	47.9	5.6	7.0	7.9	31.1	29.6	28.1	2.6	2.9	2.6	1.7	4.6	7.0
Single	392	5.04	28.0	37.2	47.6	5.6	6.6	7.9	29.2	28.4	29.4	2.6	2.7	2.8	1.5	4.3	6.8
Single	392	6.72	26.9	37.4	47.2	5.2	6.6	7.9	29.7	30.4	33.5	2.6	2.8	2.7	1.7	4.4	7.2
Split	392	1.68+1.68	28.8	37.7	48.2	5.7	7.0	8.0	29.4	27.5	28.1	2.5	3.0	2.7	1.7	4.3	6.5
Split	392	3.36+3.36	28.5	37.8	44.8	5.3	6.6	7.5	30.6	28.1	27.6	2.6	3.0	2.7	1.3	4.0	6.8
Split	392	5.04+5.04	25.1	36.6	47.0	5.0	6.6	7.8	30.9	28.0	29.7	2.6	2.7	2.7	1.2	3.6	5.5
Split	392	6.72+6.72	26.1	36.7	47.2	5.2	6.4	7.8	30.5	28.6	32.8	2.5	2.6	2.7	1.5	4.0	6.1
Single	196	3.36	28.1	38.7	47.8	5.5	6.7	7.8	29.9	29.4	29.8	2.6	2.8	2.7	1.6	3.9	6.8
Single	196	6.72	27.0	36.5	47.2	5.4	6.5	7.9	32.8	30.9	33.4	2.5	2.7	2.6	2.1	3.9	5.3
Split	196	3.36+3.36	28.6	37.0	47.4	5.6	6.4	7.9	30.9	29.7	31.4	2.5	2.8	2.6	1.4	3.4	6.4
		LSD _{0.05}	2.4	1.8	ns	0.4	0.5	ns	4.1	4.1	4.4	0.3	0.2	ns	0.6	0.9	1.3

position. Only the increased K⁺ and decreased NO₃-N were significant at the bottom stalk position of the plant. The increased K⁺, decreased Ca²⁺, Mg²⁺, P, and NO₃-N of MH-treated tobacco confirmed a previous report (48).

The Ca²⁺ content of MH-treated tobacco was 10-28% less at the top stalk position and 5-12% less at the middle stalk position compared to the hand-suckered control, but no significant difference was observed at the bottom stalk position. Lowest Ca²⁺ values were always found in leaves from the higher MH application rates, and there were no significant differences among the different application methods. When lamina and midrib content were combined for all positions, it was estimated that there was an approximate 8-10% decrease of total Ca²⁺ due to MH treatment. Because Ca²⁺ is a relatively large divalent cation with low mobility in the phloem and most of its activity is related to intermolecular linkages in the cell walls and at the plasma membrane, there is a possibility of relatively less Ca²⁺ requirement for structural functions in the leaves at the top of the plant once cell growth is inhibited by MH application. Also, the decreased Ca²⁺ could be a consequence of the antagonistic effect because there was a significant increase of K⁺ (2).

The Mg²⁺ content was affected by MH application similarly to Ca²⁺, with a 7-27% decrease in the top leaves, 3-11% decrease at

middle position, and no significant change at bottom of the plant. The estimated total decrease in Mg²⁺ in leaf tissue per plant was 6-15%. Kurvits & Kirkby (27) reported that Mg²⁺ uptake rate was strongly depressed by K⁺. Similarly, a K⁺ by Mg²⁺ interaction was reported by Hannaway et al. (19). Thus, the decreased Mg²⁺ content may be from increased K⁺ and not from a direct effect of MH.

The most abundant cation in tobacco is K⁺. It is the most important cation for tobacco burning quality and alkalinity (47), and the one most affected by application of MH. The concentration of K⁺ increased 23-39% in leaves from the top position, 9-23% from the middle position, and 16-41% from the bottom position with the addition of MH. The estimated K⁺ content for MH-treated tobacco in lamina and midrib of a whole plant was 20-40% higher than that of the hand-suckered tobacco. Similar results were reported by Seltmann & Nichols (38). As a univalent cation and charge carrier of high mobility, K⁺ is involved with osmoregulation, long-distance transport, and intracellular pH regulation (43). The role of K⁺ in cation-anion balance is also reflected in nitrate metabolism (23). Considering that the cations Ca²⁺ and Mg²⁺ decreased, it is reasonable that there was a considerable increased content of K⁺ in MH-treated plants. A portion of the increased K⁺ in MH-treated leaves could have come from the K⁺ of the applied K-salt of MH. The MH formulation

Table 5. Effects of application methods and rates of maleic hydrazide on total alkaloids at three stalk positions of air-cured burley tobacco.

Treatments			Total alkaloid at different stalk positions		
Method	Water volume	MH rate	Top	Middle	Bottom
	L ha ⁻¹	kg ha ⁻¹	----- mg g ⁻¹ -----		
Control	---	---	43.8	41.5	28.7
Single	392	1.68	37.9	35.0	24.5
Single	392	3.36	39.1	39.4	26.7
Single	392	5.04	32.5	39.9	22.8
Single	392	6.72	33.4	35.5	21.2
Split	392	1.68+1.68	36.0	38.3	22.6
Split	392	3.36+3.36	33.1	33.0	21.6
Split	392	5.04+5.04	30.3	35.3	20.9
Split	392	6.72+6.72	30.7	34.0	20.7
Single	196	3.36	39.8	37.0	24.0
Single	196	6.72	28.7	34.5	22.8
Split	196	3.36+3.36	33.2	36.9	23.3
		LSD _{0.05}	6.3	5.3	4.6

Table 6. Effects of application methods and rates of maleic hydrazide on solanesol and α -& β -diols in air-cured lamina at the top stalk position of burley tobacco.

Treatment		Chemical constituent			
Method	Water volume	MH rate	Solanesol	α -diol	β -diol
	L ha ⁻¹	kg ha ⁻¹	----- mg g ⁻¹ -----		
Control	---	---	11.9	0.08	0.04
Single	392	1.68	12.4	0.08	0.04
Single	392	3.36	11.8	0.09	0.06
Single	392	5.04	11.6	0.15	0.06
Single	392	6.72	11.3	0.14	0.05
Split	392	1.68+1.68	11.1	0.09	0.04
Split	392	3.36+3.36	10.6	0.18	0.06
Split	392	5.04+5.04	11.3	0.24	0.07
Single	392	6.72+6.72	9.7	0.18	0.06
Single	196	3.36	12.7	0.14	0.06
Single	196	6.72	10.6	0.18	0.08
Split	196	3.36+3.36	12.0	0.12	0.06
		LSD _{0.05}	ns	0.05	ns

contained 5.6% K⁺, which means that about 43.9 mg K⁺ was applied to one plant at the recommended MH application rate. Even if we assume that all the K⁺ from MH application was completely absorbed and 100% remained in the leaf, K⁺ from MH application would account for only 5% of the K⁺ increase in the leaf. This suggests that increased potassium in MH-treated tobacco was mainly a result of increased K⁺ uptake. However, because we did not measure K⁺ in the dry roots and because protein-N (calculated as total-N = NO₃-N + alkaloid-N), was increased by MH treatment, another possibility for the increased K⁺ concentration was its cycling within the plant for NO₃⁻ transport to the leaves and its subsequent association with protein-N (3).

Phosphorous content decreased significantly in lamina from the top (8-20% decline) and middle (2-13% decline) stalk positions, but no change was measured from the bottom leaves of the plant. Total leaf content of phosphorous (lamina + midrib) from whole plants decreased 3-5%. The function of phosphorous as a constituent of macromolecular structures is most prominent in nucleic acids. If cell growth were inhibited by MH, less phosphorous would be required in the younger leaves at the top of the plant and consequently less would accumulate (21).

Nitrate-N content decreased 5-48%, 5-35%, 3-28% at the top, middle, and bottom positions

of the plant, respectively. Because total-N was not affected by MH treatment and total alkaloid decreased, these data suggest that other reduced forms of N compounds (protein) must have increased. Douglass et al. (16) reported that *in vitro*, nitrate reductase activity was higher in roots and lower in leaves of MH-treated seedlings compared to control seedlings. If the same were true *in vivo*, there would be less nitrate in the leaves, as observed in this experiment. Also, Douglass et al. (15) reported that nitrate uptake rate expressed as Nitrate-N per gram dry weight of roots was equivalent for MH-treated and untreated plants. Consequently, similar total-N values were expected and changes in reduced-N forms were affected. Concentration of other anions, SO₄²⁻, Cl⁻, and NO₂⁻ in the lamina were not affected by MH treatment (data not shown).

In this study, total alkaloids of MH-treated tobacco decreased at all positions compared to the hand-suckered control (Table 5). The decreased alkaloid contents ranged 9-34%, 4-20%, and 5-29% for the top, middle, and bottom leaf positions, respectively. However, no significant effects were measured for application method at equal application rates. These results are in agreement with published studies (38).

The solanesol content (Table 6) in top leaves of MH-treated tobacco ranged 9.0-12.7 mg g⁻¹, which was not significantly different from the

hand-suckered control (11.9 mg g⁻¹). As the major terpenoid in tobacco, solanesol concentration is influenced by genotype, plant population, soil moisture, and topping (5,37). These data indicate that MH application had no appreciable influence on solanesol accumulation.

The diterpenes, α - and β -4,8,13-divatriene-1,3-diols (DVT), are present in the cuticular wax of tobacco leaves, and their degradation or pyrolytic products have distinctive flavor and aroma properties that affect tobacco quality (17,39). α -4,8,13-divatriene-1,3-diol levels increased significantly on the top leaves of MH-treated tobacco when MH application rates exceeded the recommended level (Table 6). However, β -4,8,13-divatriene-1,3-diol content on the leaves from the top stalk position ranged from 0.04-0.08 mg g⁻¹ and was not significantly affected by MH application. These observations are in contrast to those reported by Goins et al. (20) for green flue-cured tobacco. Their results were reported on mg cm⁻² basis, whereas our values are based on mg g⁻¹ basis in cured tissue. Because DVT are the most prominent terpene diols in benzene surface washes of tobacco leaves (8,34), they could be influenced by genetic factors, cultural practices, and environmental factors during the growing season (30,32). Keene & Wagner (26), Kandra & Wagner (25), and Wagner (49) suggested that CO₂ and monosaccharides produced in trichome heads or supplied by other epidermal cells were the major precursors for DVT biosynthesis, which primarily occurs in the glandular trichome heads. The inhibition of the translocation of ¹⁴C from a single leaf exposed to ¹⁴CO₂ by maleic hydrazide and the increased sugar content by MH application have been reported (6). Together these data imply that the increased DVT by high MH application rate was caused by increased biosynthesis, because direct injury or change of leaf surface property by MH were not observed (13).

In conclusion, comparison of data from all 12 treatments across two years indicates that MH application did not significantly alter dry matter accumulation and equilibrium moisture of air-cured burley tobacco. Split application and reduced water-volume application of MH significantly increased MH residues with the same level of sucker control and there was no significant change in chemical composition compared to the recommended rate of MH application. MH application decreased calcium, magnesium, phosphorus, alkaloids,

nitrate levels, whereas potassium levels were increased in cured burley tobacco. α -4,8,13-divatriene-1,3-diol content in the leaves from top stalk position of MH-treated tobacco increased significantly when MH application rates exceeded the recommended level, while β -4,8,13-divatriene-1,3-diol content and solanesol levels were not significantly changed.

ACKNOWLEDGMENTS

The investigation reported in the paper was supported by the U.S. Department of Agriculture, Agricultural Research Service under specific Cooperative Agreement No. 58-434K-0-0030 and a CORESTA Study Grant, and it is published with approval of the Director of Experiment Station (94-3-34). We wish to acknowledge Naewanna K. Dye for her help in data management and processing, George H. Childs, Jr., Novella Hounsshell, and Gordon Parmley for their assistance in chemical analyses. This paper was presented in part at the 46th Tobacco Chemists' Research Conference, Montreal, Quebec, Canada, October 1992.

LITERATURE CITED

1. Asghar, J., S. Muhammad, R. Muhammad, and S.H. Shah. Studies on the chemical control of tobacco suckers. *J. Agric. Res. Lahore*. 27:323-326. 1989.
2. Barney, P.E., L.P. Bush, and T.C. Tso. Physiologie und biochemie der tabakpflanze. 2. Physiologische storungen: Mineralstoffe. *Beitr. Tabakforsch.* 14:211-236. 1989.
3. Blevins, D.G. Role of potassium in protein metabolism in plants. Pages 413-424 *In* R.D. Munson (ed.), Potassium in Agriculture. Am. Soc. Agron., Madison, Wisc. 1985.
4. Bradstreet, R.B. The Kjeldahl Method for Organic Nitrogen. Academic Press., N. Y. 1965.
5. Burton, H.R., E. Leggett, and R.W. Phillips. Factors influencing the concentration of solanesol in burley tobacco. *Beitr. Tabakforsch.* 14: 313-320. 1989.
6. Bush, L.P., and J.L. Sims. Morphological and physiological effects of maleic hydrazide on tobacco. *Physiol. Plant.* 32:157-160. 1974.
7. Chaplin, J.F. Influences of various degrees of sucker control on flue-cured tobacco. *Tob. Sci.* 11:45-48. 1967.
8. Chang, S.Y., and C. Grunwald. Duvatrienediols in cuticular wax of burley tobacco leaves. *J. Lipid Res.* 17:7-11. 1976.

9. Cheng, S., L. Amber, and G.L. Steffens. Maleic hydrazide residue in Maryland tobacco. **Tob. Sci.** 23:29-30. 1976.
10. Crafts-Brandner, S.J., M. Collins, T.G. Sutton, and H.R. Burton. Effect of leaf maleic hydrazide concentration on yield and dry matter partitioning in burley tobacco (*Nicotiana tabacum* L.). Feasibility of using near infrared spectroscopy for rapid determination of maleic hydrazide concentration. **Field Crops Res.** (in press) 1994.
11. Crafts-Brandner, S.J., and T.G. Sutton. Effect of maleic hydrazide on photosynthesis, carbohydrate metabolism, and senescence of burley tobacco (*Nicotiana tabacum* L.). **Field Crops Res.** (in press) 1994.
12. Crutchfield, J., and H.R. Burton. Improved method for quantification of nitrite nitrogen in plant materials. **Analytical Letters** 22:555-571. 1989.
13. Currier, H.B., B.E. Day, and A.S. Crafts. Some effects of maleic hydrazide on plants. **Bot. Gaz.** 112:272-280. 1951.
14. Davis, D.L., W.O. Atkinson, and J. Smiley. Maleic hydrazide residue from air cured burley tobacco. **Crop Sci.** 14:109-112. 1974.
15. Douglass, E.A., C.T. MacKown, S.L. Gay, and L.P. Bush. Nitrate uptake and assimilation in maleic hydrazide treated tobacco. (*N. tabacum* L. cv. KY 14). **Tob. Sci.** 30:11-15. 1986.
16. Douglass, E.A., C.T. MacKown, and L.P. Bush. *In vitro* nitrate reductase activity of maleic hydrazide treated tobacco seedlings. **Tob. Sci.** 30:42-44. 1986.
17. Enzel, C.R. Terpenoid components of leaf and their relationship to smoking quality and aroma. **Rec. Adv. Tob. Sci.** 6:32-60. 1976.
18. Fiske, C.H., and Y. Subbarow. The colorimetric determination of phosphorus. **J. Biol. Chem.** 66:375. 1925.
19. Hannaway, D.B., L.P. Bush, and J.E. Leggett. Mineral composition of Kenhy tall fescue as affected by nutrient solution concentrations of Mg and K. **J. Plant Nutrition** 5:137-151. 1982.
20. Goins, G.D., D.A. Danehower, and A.R. Butler. Influence of method and degree of sucker control on the concentration of divatrienediols, yield, and quality of flue-cured tobacco. **Tob. Sci.** 37:78-83. 1993.
21. Hughes, C., and S.P. Spragg. The inhibition of mitosis by the reaction of maleic hydrazide with sulphhydryl groups. **Biochem. J.** 70:205-212. 1958.
22. Hunt, T.W., T.J. Sheets, and W.K. Collins. Maleic hydrazide residue on flue-cured tobacco. **Tob. Sci.** 21:128-130. 1977.
23. Jeschke, W.D., C.A. Atkins, and J.S. Pate. Ion circulation via phloem and xylem between root and shoot of nodulated white lupin. **J. Plant Physiol.** 117:319-330. 1985.
24. Jones, T.B., and M.H. Warner. Analysis of plant ash solutions by spark-emission spectroscopy. In E.L. Parkins (ed.), *Development in Applied Spectroscopy*. Vol. 7a. Plenum Press, N. Y. 1969.
25. Kandra, L., and G.J. Wagner. Studies on the site and mode of biosynthesis of tobacco trichome exudate components. **Arch. Biochem. Biophys.** 265:425-432. 1988.
26. Keene, C.K., and G.J. Wagner. Direct demonstration of divatrienediol biosynthesis in glandular heads of tobacco trichomes. **Plant Physiol.** 79:1026-1032. 1985.
27. Kurvits, A., and E.A. Kirkby. The uptake of nutrients by sunflower plants growing in a continuous flowing cultural system, supplied with nitrate or ammonium as nitrogen source. **Z. Pflanzenernaehr. Bodenkel.** 143:140-149. 1980.
28. Lane, J.R. Collaborative study of MH residue analyses. **J. Assoc. Offic. Analyt. Chem.** 46:261-268. 1963.
29. Lee, C.W., Y.O. Kim, B.C. Lee, and S.H. Cho. Effects of sucker control practices on growth and cured leaves of local tobacco variety Hyangcho. **Korean J. Crop Sci.** 35:53-57. 1990.
30. Leffingwell, J.C. Chemical and sensory aspects of tobacco flavor: An overview. **Rec. Adv. Tob. Sci.** 2:1-31. 1976.
31. Madsen, J.P., J.P. Bush, and S.L. Gay. Effects of curing on polyamine content of *Nicotiana tabacum* L. genotypes with different alkaloid levels. **J. Agric. Food Chem.** 33:1182-1185. 1985.
32. Nielsen, M.T. Altering flavor and aroma constituents of burley tobacco. **Tob. Sci.** 35:65-73. 1991.
33. Piro, P., and A. Vardabasso. Sucker control on burley tobacco. 9th CORESTA Congress, 9-13 October, Guanzhou, China. 1988.
34. Reid, W.W. The photochemistry of the genus *Nicotiana tabacum*. IV. Note on the diterpenes of cultivars of *N. tabacum*. **Ann. du Tabac** (Set. 2) 11:176. 1974.
35. Rodriguez, J.L. Use of maleic hydrazide in dark tobacco cv Criollo crops. I. Influences on the yield and quality. **Cienc. Teeniea Agric. Tobacco.** 12:27-31. 1989.

36. SAS institute Inc. SAS User's Guide, Version 5 Edition. SAS institute Inc., Cary, N.C. 1985.
37. Schlotzhauer, W.S., and M.J. Kasperbauer. Plant population density effects on the alkaloid, solanesol, and chlorogenic content of burley tobacco. **Tob. Sci.** 33:47-51. 1989.
38. Seltmann, H., and B.C. Nichols. Agronomic, chemical, physical, and visual characteristics of hand-suckered vs. maleic hydrazide treated flue-cured and burley tobacco. **Agron. J.** 76:375-378. 1984.
39. Severson, R.F. The cuticular chemistry of *N. tabacum*. **CORESTA Symp. Info. Bull.** 1990:34-54. 1990.
40. Severson, R.F., R.F. Arrendale, O.T. Chortyk, A.W. Johnson, D.M. Jackson, R.G. Gwynn, J.F. Chaplin, and M.G. Stephenson. Quantitation of the major cuticular components from green leaf of different tobacco types. **J. Agric. Food Chem.** 32:566-570. 1984.
41. Severson, R.F., K.L. McDuffie, R.F. Arrendale, R.G. Gwynn, and J.F. Chaplin. Rapid method for the analysis of tobacco nicotinic alkaloids. **J. Chromatogr.** 211:111-121. 1981.
42. Sheets, T.J., and L.A. Nelson. Variation of maleic hydrazide residues on flue-cured tobacco. **Tob. Sci.** 33:5-8. 1981.
43. Smith, A.F. Intracellular pH and its regulation. **Annu. Rev. Plant Physiol.** 30:289-311. 1979.
44. Steffens, G.L., D.W. Apallding, W.O. Atkinson, C.E. Bortner, L.A. Link, B.C. Nichols, H.F. Ross, H. Seltmann, and L. Shaw. Regional test with contact and systemic tobacco sucker agent. 1. Flue-cured tobacco. **Tob. Sci.** 13:113-116. 1969.
45. Steffens, G.L., D.W. Apallding, W.O. Atkinson, C.E. Bortner, L.A. Link, B.C. Nichols, H.F. Ross, H. Seltmann, and L. Shaw. Regional test with contact and systemic tobacco sucker agent. 2. Burley tobacco. **Tob. Sci.** 13:117-120. 1969.
46. Technicon Auto analyzer II, Industrial Method No. 117-71A 1972.
47. Tso, T.C. Production, physiology, and biochemistry of tobacco plant. IDEALS, Inc., Beltsville, Md. 1990.
48. U.S. Department of Agriculture. The biologic and economic assessment of maleic hydrazide. **U.S.D.A. Tech. Bull.** No. 1634. 1980.
49. Wagner, G.J. Secreting glandular trichomes: more than just hairs. **Plant Physiol.** 96:675-679. 1991.