

# EVOLUTIONARY ECOLOGY OF PROGENY SIZE IN ARTHROPODS

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■ **Abstract** Most models of optimal progeny size assume that there is a trade-off between progeny size and number, and that progeny fitness increases with increasing investment per young. We find that both assumptions are supported by empirical studies but that the trade-off is less apparent when organisms are iteroparous, use adult-acquired resources for reproduction, or provide parental care. We then review patterns of variation in progeny size among species, among populations within species, among individuals within populations, and among progeny produced by a single female. We argue that much of the variation in progeny size among species, and among populations within species, is likely due to variation in natural selection. However, few studies have manipulated progeny environments and demonstrated that the relationship between progeny size and fitness actually differs among environments, and fewer still have demonstrated why selection favors different sized progeny in different environments. We argue that much of the variation in progeny size among females within populations, and among progeny produced by a single female, is probably nonadaptive. However, some species of arthropods exhibit plasticity in progeny size in response to several environmental factors, and much of this plasticity is likely adaptive. We conclude that advances in theory have substantially outpaced empirical data. We hope that this review will stimulate researchers to examine the specific factors that result in variation in selection on progeny size within and among populations, and how this variation in selection influences the evolution of the patterns we observe.

## INTRODUCTION

Progeny size is an especially interesting life history trait because it is simultaneously a maternal and progeny character—mothers make eggs and determine egg size, but egg size can have substantial fitness effects for progeny. Thus, progeny size is subject to selection in both the parental and progeny generations. This selection often varies in direction and/or magnitude among generations (parental versus offspring), among environments, and even among siblings within

a family, such that understanding the factors that influence the evolution of progeny size can become quite a challenge. In this review, we focus on understanding the causes and consequences of egg and progeny size variation in arthropods. Although most arthropods lay eggs, many crustaceans brood their eggs and studies of progeny size in crustaceans measure progeny after eggs hatch and are released by the parent. We thus use the phrases “egg size” or “progeny size” interchangeably. We also acknowledge that eggs and progeny often vary in ways other than size (e.g. egg composition) and that this variation may be ecologically and evolutionarily as important as variation in size (17). However, due to space constraints we limit our discussion to progeny size.

We begin with a brief discussion of the optimality model developed by Smith & Fretwell that laid the foundation for how we think about the evolution of progeny size (197). We focus on empirical studies that examine the two primary assumptions of this model. Next, we explore patterns of variation in progeny size among species and among populations within species. Lastly, we review the sources of variation in progeny size within populations and discuss proposed explanations for this variation.

## CONCEPTUAL FRAMEWORK

Christopher Smith and Steven Fretwell (197) offered the first mathematical analysis of optimal progeny size. They asked “what size progeny should a female produce to maximize her total number of grandprogeny?” The number of grandprogeny a female will produce depends on both the number of progeny she produces and the fitness of those progeny. To model this, Smith & Fretwell started with two assumptions: (a) progeny fitness ( $W_{\text{Young}}$ ) increases with increasing parental investment per offspring ( $I_{\text{Young}}$ ) (i.e. larger progeny will have higher fitness), and (b) for any fixed amount of parental investment into reproduction ( $I_{\text{Total}}$ ), a female can produce  $N = I_{\text{Total}} / I_{\text{Young}}$  progeny. In other words, there is a trade-off between the number of progeny a female can make and the amount of resources allocated to each of those progeny. If a female makes larger progeny,  $I_{\text{Young}}$  increases and  $N$  decreases. To increase  $N$ , a female must either decrease  $I_{\text{Young}}$  or increase  $I_{\text{Total}}$ . Smith & Fretwell assumed that  $I_{\text{Total}}$  is a constant. Maternal fitness,  $W_{\text{Parent}} = N \times (W_{\text{Young}}) = (I_{\text{Total}} / I_{\text{Young}}) \times (W_{\text{Young}})$ , the product of the number of progeny that she produces times the fitness of each of those progeny. The value of  $I_{\text{Young}}$  that results in the highest parental fitness is the value that maximizes  $(I_{\text{Total}} / I_{\text{Young}}) \times (W_{\text{Young}})$ . Maternal fitness thus increases as  $W_{\text{Young}}$  increases, but also increases as  $(I_{\text{Total}} / I_{\text{Young}})$  increases (i.e. fecundity increases). The constraint here is that for any fixed amount of resources ( $I_{\text{Total}}$ ), females can increase  $W_{\text{Young}}$  only by increasing  $I_{\text{Young}}$ , which necessarily results in a decrease in fecundity ( $I_{\text{Total}} / I_{\text{Young}}$ ).

This model illustrates three points that have become the subject of much empirical and theoretical exploration. First, for any fixed parental allocation to repro-

duction, progeny size is under balancing selection; large progeny are favored because  $W_{\text{Young}}$  increases as  $I_{\text{Young}}$  increases, and small progeny are favored because  $N$  increases as  $I_{\text{Young}}$  decreases. Second, there is a conflict of interest between parents and their progeny. Because progeny fitness ( $W_{\text{Young}}$ ) increases with increasing investment per progeny ( $I_{\text{Young}}$ ), the value of  $I_{\text{Young}}$  that maximizes progeny fitness is larger than the value that maximizes parental fitness. Third, any environmental variable that affects the relationship between investment per progeny and progeny fitness (i.e. between  $I_{\text{Young}}$  and  $W_{\text{Young}}$ ) can result in a change in the optimal progeny size and thus a change in the size of progeny that should evolve in a population. The first and third of these points will be discussed in this paper. The consequences of conflicts of interest between parents and their offspring has been reviewed extensively elsewhere and will not be discussed here.

Since the original development of the Smith-Fretwell model, more complex models have been developed to examine optimal progeny size under more specific conditions (46, 182). It is not the objective of this paper to review the various models and their specific assumptions (see 17). However, most of these models start with the same basic assumptions that Smith & Fretwell started with, that (a) there is a trade-off between progeny size and number, and (b) progeny fitness ( $W_{\text{Young}}$ ) increases with increasing parental investment per offspring. We thus focus first on these two assumptions.

### Trade-Offs Between Progeny Size and Number

The concept of trade-offs is an integral part of life history theory (182). If an individual has a fixed amount of resources available, those resources can be divided into three basic functions—growth, somatic maintenance, or reproduction. Resources directed to reproduction can subsequently be divided into either many small progeny or a few larger progeny. Thus, for a fixed amount of resources allocated to reproduction it necessarily follows that there is a trade-off between the number and size of progeny

Phenotypic correlations between egg size and number (a) among species (18, 19, 33, 42, 65, 84, 86, 127, 129, 139, 143, 175, 177, 193, 204, 217), (b) among populations within species (2, 56, 127, 230) and (c) among individuals within populations (Table 1) generally indicate a trade-off between egg size and number. Most of these studies examine only phenotypic correlations between egg size and number, but a genetically based trade-off has been demonstrated for *Daphnia* (57, 140).

In general, trade-offs have been detected in most studies of relatively semelparous arthropods that use larval-acquired resources for egg production and exhibit no parental care (Table 1). In studies of more complex systems (especially vertebrates), in which females are iteroparous, use adult-acquired resources for reproduction (e.g. shrimp, mosquitoes), or exhibit parental care (e.g. birds), a trade-off has been more difficult to demonstrate (87), leading some authors to suggest that such a trade-off is not universal (e.g. 17). Failure to demonstrate

**TABLE 1** Evidence for (or against) a trade-off between egg/progeny size and number, based on variation among females within a population (number of species)

Taxon	Trade-off	No Trade-off	Reference (trade-off)	Reference (no trade-off)
Crustacea (cladoceran)	7	1	15 <sup>2</sup> , 22, 45, 87 <sup>1</sup> , 140, 153, 213	15 <sup>2</sup>
Crustacea (copepod)	2	0	2, 93	
Crustacea (shrimp)	0	4		42
Crustacea (isopod)	2	1	135, 230	50
Orthoptera	5	0	39, 68 <sup>3</sup>	
Heteroptera	3	0	132, 154, 198	
Lepidoptera	3	3	66 <sup>2</sup> , 136, 180	16, 26, 146 <sup>4</sup>
Coleoptera	3	0	81, 94, 219	
Diptera	3	2	4 <sup>2,5</sup> , 138	4 <sup>2</sup> , 29

<sup>1</sup>Varied with age of the female (iteroparous organism)

<sup>2</sup>Based on variation in egg size through the season

<sup>3</sup>Confounded with maternal age

<sup>4</sup>Did not control for female size

<sup>5</sup>Varied among studies

trade-offs probably has less to do with their absence than with the complexity of the system. For a trade-off between egg size and number to be evident, we must assume that the quantity of resources allocated to reproduction ( $I_{\text{Total}}$ ) is constant. Yet  $I_{\text{Total}}$  is often not constant. For example, variation in larval growth can produce substantial variation in body size at maturation, which generally corresponds closely to total reproductive effort (within a population). Thus, larger individuals generally lay both more and larger eggs, leading to a positive correlation between egg size and number. In this case, the relationship between egg size and number will be negative only when body size is controlled (e.g. 16, 40, 81, 154).

Other sources of variation in reproductive effort are less easily quantified and controlled, including variation in adult feeding rates, the proportion of adult-acquired resources allocated to reproduction, degree of parental care, etc. Our conclusion is that the assumption of a trade-off between egg size and number is generally supported by empirical studies in arthropods; studies that have failed to detect such a trade-off have generally been on animals in which there may be substantial variation in reproductive effort obscuring the patterns.

We suggest that rather than testing for the presence or absence of trade-offs between progeny size and number, future research should focus on two general issues. First, we know of little empirical data on the shape of the relationship between progeny size and number, although theoretical predictions often depend on an assumed shape (but see 40a). Smith & Fretwell (197) originally proposed that the number of offspring produced by a female is a simple function of  $I_{\text{Total}}$  and  $I_{\text{Young}}$ ; the female can produce  $I_{\text{Total}}/I_{\text{Young}}$  progeny. However, it is likely that,

due to inefficiencies in resource allocation, allocating  $I_{\text{Total}}$  resources to reproduction does not allow for the production of  $I_{\text{Total}} / I_{\text{Young}}$  progeny of size  $I_{\text{Young}}$ ; dividing resources among progeny may not be as simple as dividing a pie into pieces. Second, we have little understanding of how changes in reproductive effort affect the relationship between progeny size, progeny number, and maternal fitness (230a). Reproductive effort may evolve as a result of changes in female survival probabilities, changes in resource availability, or due to selection on progeny size or fecundity (e.g. 182). Smith & Fretwell (197) and most models since have assumed that total reproductive effort is constant (but see 230a). More theoretical and empirical exploration of these two issues is needed.

### Fitness Consequences of Progeny Size

Many studies have examined the relationship between egg size and fitness components of progeny. They often demonstrate that smaller eggs hatch more quickly (7, 72) or are brooded for a shorter time (231), but are less likely to hatch (7, 48, 69, 70, 154; but see 93, 146, 214). Progeny hatching from smaller eggs tend to be smaller hatchlings (7, 9, 27, 31, 38, 46a, 92–94, 130, 132, 161, 178, 187, 225) that grow into smaller-than-normal later instars (118, 132, 135, 219) and have lower juvenile survivorship (27, 36, 38, 40, 70, 78, 81, 110, 119, 168, 214, 219; but see 49, 72, 202).

Smaller-than-average young have three developmental options: (a) mature at a smaller-than-average size (27, 36, 38, 79, 90, 109, 118, 130, 154, 202; but see 9, 49, 179, 180), (b) extend development to fully or partially compensate for their small starting size (7, 27, 56, 70, 72, 73, 76, 94, 101, 137, 154, 179, 184, 190, 202, 214; but see 9, 118, 219, 229), or (c) increase their rate of growth to mature at a normal size. Most arthropods exhibit some degree of developmental plasticity by which progeny partially compensate for their small hatchling/birth size by extending development time (72, 73). Few studies have examined the influence of juvenile size on growth rates in arthropods (but see 7). Progeny hatching from larger eggs can often better withstand environmental stresses such as larval competition (7), starvation (38, 89, 145, 199, 212), desiccation (201), oxygen stress (97), cold stress (36, 105), nutritional stress (27, 74, 78, 219), and environmental toxins (62). Some studies have failed to detect fitness advantages of hatching from large eggs; most of these studies have raised progeny in high-quality environments (e.g. 118, 228, 229), suggesting that selection is generally weak in high-quality environments but favors larger eggs in lower-quality environments (74, 189).

Most of the studies cited here are correlational studies that confound relationships between egg size and progeny fitness with genetic correlations between morphological and life history characters (194). For example, larger females generally lay larger eggs and produce progeny that mature at a larger size (because body size is generally heritable) such that there is a positive correlation between egg size and progeny size at maturation (71). Experimental approaches were thus developed to study the consequences of, and selection on, egg size variation (194).

By manipulating egg size physically or physiologically we can quantify effects of egg size variation on progeny fitness. A few studies have manipulated egg size in invertebrates (61, 100, 195) including one insect species (72). They have generally demonstrated that progeny hatching from larger eggs do indeed have higher fitness or improved performance (but see 99).

### Time Limitation, Parental Care, Clutch Size, and Constraints on Progeny Size

Smith & Fretwell assumed that all eggs of size  $I_{\text{young}}$  have the same influence on a female's fitness such that maternal fitness is the product of the average fitness of her offspring times the number of progeny produced (197). However, this model assumes that females can actually lay all of their matured eggs. In many parasitic insects (e.g. herbivores and parasitoids) females may be incapable of finding enough hosts to lay all their eggs, relaxing selection for increased fecundity (177, 228) and potentially shifting the optimal egg size to a larger value than predicted by the Smith-Fretwell model (86). Thus, shifts in the abundance of hosts may result in a change in optimal egg size, even without changes in the relationship between egg size and progeny fitness (183).

The Smith-Fretwell model also assumes that maternal fecundity influences progeny fitness only by affecting progeny size. However, for organisms that exhibit parental care, large clutches may be less easily tended/defended than smaller clutches, such that progeny survivorship decreases with increasing maternal fecundity even if progeny size is constant. Similarly, progeny within larger clutches may experience increased competition or conflict that decreases progeny fitness (169). Thus, both parental care and sibling competition can select against large clutches (but see 193), resulting in a change in optimal progeny size without a change in the relationship between progeny size and progeny fitness.

Finally, there may be morphological and physiological constraints on the ability of females to make especially large or small eggs. For example, the necessity for progeny to fit into the brood pouch of a female may constrain the evolution of large progeny in *Daphnia* (181), even when large progeny are favored by environmental conditions. Unfortunately, although some physical and physiological constraints on progeny size have been studied in vertebrates (17, 46, 182), constraints have been little examined in arthropods.

## VARIATION IN PROGENY SIZE AMONG SPECIES AND AMONG POPULATIONS WITHIN SPECIES

### Selection on Progeny Size Varies Across Space and Time

When environmental conditions vary, the relationship between progeny size and progeny fitness is likely to vary, resulting in different optimal progeny sizes in different environments. However, few studies have manipulated progeny envi-

ronments and quantified the relationship between egg size and progeny growth or survival in each environment. These studies have demonstrated that selection on egg size varies across environments (27, 40, 74, 78, 154, 180). In general, it appears that the fitness difference between progeny hatching from large vs. small eggs is greatest in lower quality or more stressful environments (27, 74, 78).

Climatic conditions vary substantially across space and time and may result in substantial variation in selection on progeny size (6, 180). In some insects, selection on egg size may depend on whether progeny need to overwinter before hatching (39, 66, 105, 125, 131, 178). Variation in season length or the amount of time left before winter may impose variable selection on development time, in which selection for rapid development of progeny produced late in the season (or progeny living in areas with short or cool summers) favors progeny hatching from large eggs (8, 169).

Selection on egg size can vary with the depth at which eggs are laid in the soil (crickets, 39) and the host species upon which eggs are laid (herbivores, 27, 74, 78, 161). Population density can affect the amount of competition for food that progeny will encounter, which may affect selection on egg size (169). At low population densities, sperm limitation becomes important for free-spawning arthropods, and selection may favor the evolution of large eggs that are more likely to be fertilized (206). Egg size may affect the ability and tendency of larvae to disperse (9, 16), such that variation in the need to disperse will influence selection on egg size.

Size-specific predation may represent an important source of selection on progeny size (128) either by influencing the demographic environment or because smaller progeny may be less susceptible to visual predators (30, 123, 139). For terrestrial insects size-selective egg predators and parasites impose selection on egg size that will vary with predation intensity. When predation on immature stages is high, selection may favor progeny that spend less time as juveniles, thus favoring progeny that start life larger (188). Egg size of predators may be constrained by the minimum size at which hatchlings can capture prey (204) such that selection intensity varies with prey size (1, 129; but see 204). Also, larger eggs may be favored at low prey densities to protect against periods of starvation encountered after egg hatch (129, 209).

Selection may also vary among progeny produced by a single female due to small-scale environmental variation. For example, selection on egg size varies among trees in the seed beetle *Stator limbatus*. Theoretical models predict that increased variability in selection on progeny size within populations will result in selection for larger progeny than predicted by the Smith-Fretwell model (67). Alternatively, variation in selection can result in the evolution of increased variance in progeny size (115, 174) or plasticity in progeny size (46, 182).

### Variation in Progeny Size Among Species

Within genera or families, females of larger species generally lay larger eggs than females of smaller species (3, 18, 19, 33, 84–86, 90, 108, 139, 176, 204, 223), suggesting morphological constraints on egg size. However, in many taxa vari-

ation in female body size does not explain among-species variation in egg size (28, 64, 65, 129, 175, 176, 211, 228) and, even when female body size is correlated with egg size, there is generally substantial variation around the regression line (85, 124, 181, 193). Although females of larger-bodied species generally lay larger eggs, they often allocate a smaller proportion of their resources to each egg (3, 18, 85, 139, 147, 181; but see 223).

Few studies have examined the causes of variation in egg size among species. In many crustaceans, marine- and brackish-water species differ from inland species in both the size and number of eggs laid (98, 144). Higher-latitude shrimp (42) and satyrid (84) species generally lay larger eggs than lower-latitude species, while higher-latitude cladoceran species on average lay smaller eggs (175; but see 176), suggesting climate-mediated adaptive differentiation. Mode of parasitism explains some of the interspecific variation in egg size among parasitic cladocerans (175). Variation in relative egg size among species of cladocerans (in which smaller species produce proportionately larger eggs; 181) may be due in part to size-specific predation on progeny (139); small cladoceran species may minimize juvenile mortality by producing relatively larger progeny that quickly attain adult body size and reproduce before they are subject to predation. However, this pattern of negative allometry is observed in many other arthropods for which size-selective predation is not likely a source of selection (see above).

Marine arthropods with planktotrophic larvae produce smaller eggs than species with direct developing larvae (41, 100). Host plant toughness may influence the evolution of skipper (hesperiid) egg sizes; species that oviposit on hosts with tougher leaves lay larger eggs (160; see also 178; but see 85). In stored-products insects, the relationship between body size and egg size is different for semelparous versus iteroparous species; semelparous insects produce smaller eggs (relative to body size) and the slope of the relationship between egg size and body size (among species) is less steep, but the explanation for this pattern is unclear (108).

In ponerine ants, selection for large colony size appears to explain among-species variation in egg size (small eggs in species that produce large colonies; 217). Egg size of carabids varies among species according to prey type (219), and in some herbivore taxa specialist feeders lay larger eggs than generalist feeders (64, 86; but see 178), possibly as a result of relaxed selection on fecundity due to difficulty finding enough hosts (183). In Lepidoptera, species that overwinter as eggs tend to lay larger eggs than species that overwinter in other stages (178; but see 85).

## Variation in Progeny Size Among Populations Within Species

Within species, females from larger-bodied populations tend to lay larger eggs (13, 228). However, variation in body size alone cannot account for the substantial geographic variation observed in many arthropods (43, 55, 83, 199, 230). Egg size often follows a cline in latitude (6, 13, 20, 43, 44, 91, 103), altitude (13, 98,



158), or, for crustaceans, habitat predictability (permanent versus temporary pools; 13, 158), from coastal to inland waterways (149, 150, 163, 165, 220) or from deep-sea benthic to shallow coastal waters (165). Some of these clines are known to be genetically based (6, 103, 151). Most cannot be explained entirely by clines in female body size (6, 13, 44, 103).

Most intraspecific latitudinal clines go from smaller eggs produced at lower latitudes to larger eggs at higher latitudes (6, 13, 43, 44, 91, 99, 103; see also 2, 21, 199), although some insects exhibit the opposite pattern (3, 20, 83). The commonness of these latitudinal clines is often interpreted as evidence that large eggs are selectively favored at low temperatures. However, environmental effects of temperature on egg size often mimic the geographic clines observed in nature (larger eggs at lower temperatures; see below).

Only one study (by Azevedo et al.) has experimentally demonstrated that eggs evolve to be larger when populations are reared at low temperatures; *Drosophila* maintained for nine years at 16.5° C evolved larger egg sizes than flies maintained at 25° C (6). However, it is unclear why larger eggs are favored at lower temperatures (6, 63, 180; see below). For some insects, short growing seasons may constrain fecundity of females in northern latitudes, relaxing selection for small eggs (228), but this hypothesis does not explain the results of Azevedo et al (6). Egg size clines in aquatic crustaceans have been argued to be due to variation in water temperature (165) or salinity (98). However, clines vary substantially in form and direction among species (148, 150, 151, 163, 165), suggesting alternative explanations.

Other explanations have been proposed to account for latitudinal clines. For example, food availability in polar environments may select for relatively K-selected life history strategies, including the production of a small number of highly competitive progeny, a pattern typical of polar benthic organisms (42). Variation in food availability has also been proposed to explain coastal-to-inland clines (149) and altitudinal clines (98) in crustacean egg size.

For many arthropods, variation in egg size among populations does not appear to be clinal. In many crustaceans, egg size varies among lakes or bays (12, 25, 56, 127, 142, 155, 205). In some herbivores, egg size varies among populations using different host plants (27, 78). Variation in egg size among populations may be due to variation in the need to resist desiccation (201; but see 200) or compete with conspecifics for food (169). Each of these studies suggests adaptive differentiation of egg size among populations, but in most cases the explanation for the differentiation is unclear or untested.

## VARIATION IN PROGENY SIZE AMONG FEMALES WITHIN POPULATIONS

### Female Size

Within populations, larger females tend to lay larger eggs (Table 2), suggesting some morphological constraints on egg size. However, there are many exceptions to this pattern: In some butterflies larger females lay smaller eggs, and in most

**TABLE 2** Phenotypic correlations between maternal size and egg/progeny size, within populations (number of species)

Taxon	Posi- tive	Nega- tive	Variable or No Relationship	Reference
Crustacea (cladoceran)	9	0	0	Positive (22, 87, 88, 90, 97, 123, 130, 153, 172, 181)
Crustacea (copepod)	1	0	0	Positive (101, 155)
Crustacea (shrimp)	1	0	6 <sup>1</sup> , 1 <sup>2</sup>	Positive (220); Variable (12, 13); None (158, 165)
Crustacea (isopod)	2	0	5 <sup>1</sup> , 1 <sup>2</sup> , 1 <sup>5</sup>	Positive (211, 230); Variable (43); None (49, 50, 135, 211)
Crustacea (lobster)	0	0	1 <sup>1</sup>	None (203)
Ephemeroptera	0	0	1 <sup>1</sup>	None (46a)
Orthoptera	4	0	6 <sup>1</sup>	Positive (38, 131); None (35, 40, 68)
Heteroptera	5	0	2 <sup>1</sup> , 1 <sup>4</sup>	Positive (52, 53, 132, 154, 162); Variable (208); None (110, 199)
Lepidoptera	6	2	3 <sup>1</sup>	Positive (16, 26, 107, 116, 117, 146, 164); Negative (102, 113); None (21, 180, 207)
Coleoptera	5	0	1 <sup>1</sup> , 1 <sup>5</sup>	Positive (69, 71, 94, 112, 122, 134, 168); Variable (81); None (114)
Diptera	2	0	5 <sup>1</sup>	Positive (202, 224); None (29, 65, 99, 138)
Hymenoptera	7	0	0	Positive (133, 166)

<sup>1</sup>No relationship<sup>2</sup>Varied among populations<sup>3</sup>Varied among clones<sup>4</sup>Varied among wing morphs<sup>5</sup>Varied among studies

isopods and orthopterans there is no relationship between progeny size and female size (Table 2). Even when the relationship between female size and progeny size is positive, it is generally weak (e.g. 81), and larger females generally allocate a smaller proportion of their resources to each egg. Numerous authors have advanced adaptive explanations for why egg size should increase with female size within populations (24, 46, 152). We suggest that an equally interesting question is why (physiologically and evolutionarily) the proportion of a female's resources allocated to each egg generally decreases with increasing body size. It is likely that the degree to which egg size varies with body size is in part influenced by where resources come from during egg maturation. For insects that obtain most resources for egg production from adult feeding, the size of eggs laid by females may be more dependent on female diet than female size (202), while the reverse may be true if mostly larval-derived resources are used.

## Maternal Diet/Food Availability

Maternal diet influences egg size in many arthropods. Generally, unfed or food-stressed females lay smaller eggs than well-fed females (28, 69, 104, 117, 124, 141, 159, 202, 219). However, there are many examples in which maternal diet does not affect egg size (63, 96, 99, 107), has only a small effect on egg size (106), or affects egg size only when females are extremely food stressed (104).

Theoretical models generally predict that, as food availability decreases, and thus progeny mortality increases, females should shift to laying larger eggs (46, 189). In some crustaceans females produce larger progeny at low food concentrations (*Daphnia*: 23, 24, 34, 58, 87–89, 92, 153, 173, 179; *Euterpina*: 93; and one isopod: 32), although progeny size may decrease at very low food levels (22, 213, 215). This increased progeny size often results in higher survivorship under food stress (89; references in 22). In some *Daphnia* the response to food concentration varies among clones (60, 87, 88, 213), indicating the potential for adaptive evolution of egg size plasticity. The environmental cues to which females respond, and the physiological mechanisms by which they respond, are still unknown (88).

## Oviposition Host

Some insects modify egg size in response to the host plant upon which they mature eggs (81, 136). For the seed beetle *Stator limbatus*, hosts vary in the degree to which their seeds are defended against larvae. On well-defended hosts larval mortality is high and selection favors females that lay large eggs (74, 78, 81, 82). On undefended hosts larval mortality is low and selection favors females that lay small eggs (and thus have high fecundity; 78, 81). Apparently in response to this variation in selection, females have evolved egg size plasticity—they lay large eggs on seeds of the well-defended host and small eggs on seeds of undefended hosts (81, 82). The degree of plasticity exhibited by females is genetically variable within populations (75). Interestingly, this plasticity appears to mediate a diet shift by *S. limbatus* onto an exotic legume (80).

## Maternal Density

Females reared at high densities often lay eggs that are smaller than those of females reared at low density (73, 76, 79, 154; but see 65, 185), likely due to effects of competition on female size or nutritional status. In some cladocerans, females respond to increased population density by producing larger progeny (173), which can better tolerate periods of starvation (45, 89) and may compete better for food. This plasticity may be mediated by sensitivity to the chemical (e.g. waste products) or physical cues emitted by other individuals (34), or by effects of density on food availability. Similar egg size plasticity in response to perceived larval competition has been reported for a seed beetle (122).

## Paternal Effects

In most arthropods, nutrients and other substances are transferred to females during mating and provide a pool of resources for females to use during egg maturation (216). These contributions may affect female egg size by being incorporated directly into eggs or by changing female energy budgets. Many studies have examined how male contributions affect female survivorship or fecundity (216), but few have examined whether they affect the size or composition of eggs. Female insects sometimes lay larger eggs when they receive more (95, 96, 164, 192) or larger spermatophores (96), although the effect is sometimes seen only late in a female's life (69, 222). Some insects lay larger eggs when they mate with larger males (154, 225), possibly as a result of paternal investment or because males manipulate female allocation to the eggs they have fertilized (167).

Other studies have failed to find effects of female mating frequency (35, 207), spermatophore size, or male size (76, 79, 186) on egg size. Some authors have suggested that effects of male-derived nutrients on female reproduction may be detectable only when females are food stressed (but see 69). The relative influence of male-derived nutrients on egg size is still unclear.

## Rearing and Oviposition Temperature

Many studies show that females lay larger eggs when reared (104) or ovipositing (4, 5, 63, 101) at lower temperatures (232), although some arthropods lay larger eggs when reared at intermediate temperatures (10), lay larger eggs at high temperatures (110), exhibit variable responses to temperature depending on other environmental conditions (e.g. food availability, 153), or show no response to temperature (221). Unfortunately, many studies do not distinguish between the effects of rearing versus oviposition temperature (10, 30, 47, 172, 181, 187). Interestingly, the temperature at which *D. melanogaster* males are reared affects the size of eggs laid by their daughters (47), but the mechanism and adaptive significance for this environmentally-based paternal effect is unknown.

The rate of oocyte production relative to the rate of oocyte growth (vitellogenesis) may change with temperature, affecting both the size and number of eggs (63). If so, the temperature at which vitellogenesis occurs should affect egg size (218), and an increase in egg size should be accompanied by a decrease in fecundity, as generally observed. The size of a female's fat body may be affected by temperature (but see 63) and may in turn affect the rate of vitellogenin uptake. This hypothesis predicts that only temperatures experienced during fat deposition (prior to oviposition) should affect egg size, and that both egg size and fecundity should be affected similarly by temperature (both increase or decrease), neither of which is generally observed.

Temperatures experienced by adults may affect the metabolic rate of females (4): If low temperature reduces the cost of somatic maintenance, a greater proportion of the female's resources may be shunted to vitellogenesis. Some arthro-

pods mature at a larger body size when reared at lower temperature (e.g. most cladocerans; 156), potentially resulting in an increase in egg size (e.g. Table 2). However, females of many species respond to oviposition temperature independent of rearing temperature (4, 5, 63, 101), indicating that a change in body size is not a general explanation (see also 172).

Other arthropods delay oviposition at lower temperatures (e.g. *Drosophila*; 111), potentially resulting in increased vitellogenesis. This hypothesis predicts that egg size should vary with manipulations of oviposition rate independent of temperature. In some insects, delaying oviposition or changing oviposition rate affects egg size (219; references in 4), but in other insects, females forced to delay oviposition do not lay larger eggs (e.g. 81). Also, in some insects the production of larger eggs does not result in an increase in the period of oogenesis or delayed oviposition (54).

Increasing egg size at low temperatures may represent an adaptive response to temperature (232). For example, at lower temperatures growth is slower, so selection may favor the production of larger progeny that mature sooner, reducing their exposure to sources of mortality (232) or simply decreasing generation time (91, 172). Few other adaptive hypotheses have been proposed (see 181, 232) and none have been tested.

### Seasonal Variation

In many arthropods, progeny size varies throughout the year (4, 5, 28, 32, 46a, 50, 63, 66, 110, 125, 161). In some cases this is due to aging of females in the population and corresponding changes in egg size (Table 3). However, seasonal variation sometimes reflects variation among generations (27, 28, 161), and maternal age cannot explain some of the patterns observed within generations (46a, 63).

Most species of crustaceans that exhibit seasonal variation in progeny size produce larger progeny in winter (15, 30, 33, 93, 123, 130, 155, 187, 196, 231), although some species produce larger progeny in summer (15), and others show some other seasonal pattern (15). In some species, females may be responding primarily to temperature, but it is unclear whether the responses are adaptations to temperature itself, non-adaptive physiological responses to temperature, or whether temperature is used as a cue to predict some other environmental condition. However, temperature cannot explain the seasonal pattern observed in some other species (93, 123, 155).

Seasonal variation in progeny size often reflects variation in female size (30, 123, 155) although it is unclear whether this reflects a cause-and-effect relationship or whether body size and egg size are influenced by the same external factors. In some crustaceans seasonal differences in body size cannot explain all of the variation in egg size (230). Seasonal changes in cladoceran progeny size often correspond to changes in predator abundance, suggesting an adaptive response to variation in size-specific predation (22, 30, 123, 130). Alternatively, seasonal variation may reflect a plastic response to variation in food availability (91; but

**TABLE 3** Change in egg or progeny size as females age (number of species)

Taxon	Increase	Decrease	Varied or no change	Reference
Crustacea (cladoceran)	2	0	0	Increase (14, 24, 62, 87, 130, 153)
Crustacea (isopod)	1	0	0	Increase (32)
Orthoptera	2	4	1 <sup>1</sup> , 3 <sup>2</sup>	Increase (35, 131); Decrease (40, 68, 131); No change (131); Varied (37, 40)
Heteroptera	6	2	2 <sup>2,3</sup>	Increase (51, 52, 120, 154, 162); Decrease (110, 132); Varied (155a, 198)
Lepidoptera	0	20	2 <sup>1</sup> , 1 <sup>4</sup> , 1 <sup>5</sup> , 1 <sup>6</sup>	Decrease (26, 28, 36, 83, 102, 107, 113, 116, 118, 119, 126, 136, 137, 164, 180, 185, 207, 210, 226, 227, 229); No change (16, 146); Varied (28, 106, 157, 159)
Coleoptera	0	2	1 <sup>4</sup>	Decrease (69, 77, 219, 222); Varied (81)
Diptera	0	0	1 <sup>1</sup>	No change (221)

<sup>1</sup>No change<sup>2</sup>Varied among females<sup>3</sup>Varied among morphs<sup>4</sup>Varied among treatments<sup>5</sup>Varied among seasons<sup>6</sup>Varied among studies

see 30) or clonal replacement, in which natural selection results in the replacement of large-egg clones with small-egg clones in warmer seasons (130).

In terrestrial arthropods, no consistent seasonal patterns are apparent, so temperature is not a general explanation for seasonal variation. In some insects, seasonal changes in body size correspond to changes in egg size (27). Eggs laid by second-generation females of a tortricid moth (which enter diapause) may be larger to ensure overwinter survivorship of diapausing eggs (66; see also 125, 131). In some herbivores selection on egg size varies among host plants and a seasonal change in egg size may be an adaptation to changes in host plants availability (27, 161). In isopods, seasonal variation in egg size may reflect a response to food availability (32).

## Responses to Predation Risk

Some cladocerans exhibit plasticity in progeny size in response to predator-associated chemical cues (97, 179). Populations of *Daphnia magna*, and clones within populations, vary in their responses to fish kairomones, with clones from lakes with fish generally more sensitive (25). Other arthropods may also respond to predation risk by varying egg size. For example, females of the shield bug

*Elasmucha ferrugata* lay smaller eggs at the periphery of their clutch where the eggs are most susceptible to predation (145) and thus have lower reproductive value.

### Other Environmental Sources of Variation

Many insects exhibit complex polymorphisms in suites of morphological and life history characters, and egg size often differs substantially among morphs. For example, macropterous individuals of both *Lygaeus equestris* and *Orgyia thyelina* lay smaller (and more) eggs than brachypterous individuals, possibly as a result of selection for rapid population increase (and thus high fecundity) on females that colonize new habitats, and selection for producing large, competitive progeny on females that stay in established populations (125, 198). However, the opposite pattern is observed in *Jadera aeola* (208). Alatae of polymorphic aphids generally produce smaller offspring than apterae (52, 162). This pattern has been attributed to competition between gonads and flight muscles for limited resources (162). Obligate asexual clones of *Daphnia* produce larger progeny than sexual clones (232), but parthenogenetic eggs do not differ in size from fertilized eggs in a stick insect (37).

### Genetic Variation

There are surprisingly few data available on genetic variation in egg size within populations of arthropods. Estimates of the heritability of egg size, and its genetic correlations with other life history traits, are even fewer. In two seed beetles, egg size is highly heritable (range of  $h^2 = 0.22$  to  $0.91$ ;  $h^2$  varies among hosts and populations; 71, 75). Likewise, variation in egg size is heritable in spruce budworm (104). Comparisons of clones of *Daphnia* indicate substantial genetic variation in both progeny size (25, 56, 59, 87, 140) and egg size plasticity (25, 60, 87) within populations. Laboratory selection experiments have also demonstrated that egg size is heritable (7, 38, 170, 186a, 221) and is genetically correlated with body size. Selection on other life history characters, such as development rate (8) and resistance to desiccation (200) have also resulted in the evolution of egg size, indicating genetic correlations between egg size and these traits.

Little is understood about the genetic basis of among-species or among population differences in egg size. Crosses among strains of *D. melanogaster* (221) suggest that at least one autosomal and one sex-linked gene affect the variation in egg size (among strains). Egg size variation among species of *Choristoneura* (36) and among strains of silkworms (121) is also partially sex linked.

### VARIATION AMONG EGGS AND PROGENY PRODUCED BY A SINGLE FEMALE

For many arthropods, the variation in size among progeny produced by a single female may be as large as or larger than the variation among females within a

population. Much of this variation is an effect of maternal age (below), but in many arthropods there is substantial variation in egg size within individual clutches of eggs. Variation in egg size within and among clutches may be selected for as a diversified bet-hedging strategy to minimize variation in fitness (174; see 67a for a related adaptive explanation). Alternatively, physiological limitations in the ability to make identically sized eggs may explain much of the egg size variation within clutches.

### Female Age/Egg Order

Most life-history models posit the production of uniform-sized progeny throughout a female's life (e.g. 197). In most arthropods, however, progeny size decreases with maternal age (Table 3), although an increase is commonly observed in orthopterans and heteropterans (Table 3). Only a few insects exhibit no change in egg size (16, 40, 146). An increase in progeny size is commonly observed in cladocerans (24; Table 3), but this is because females continue to grow after beginning reproduction; the ratio of progeny size to maternal size actually decreases with age (153). In some insects, maternal age effects are not observable until females near their last clutch (157). The variance in egg size sometimes also increases as mothers age (171), but too few studies present estimates of variance to allow generalization.

The effect of age often varies substantially among females (37, 102, 131, 119, 228, 229) but the degree to which variation among females reflects genetic differences is unclear. At least some of the variation is environmentally based. For example, the direction or magnitude of the maternal age effect can differ among host plants (81), between macropterous and brachypterous bugs (198), and between alatae and apterae of aphids (51). The maternal effect may also vary with maternal diet (28; but see 107), with the decrease generally steepest for food-stressed mothers (69, 159) or for females that had been food stressed as larvae (102).

A decrease in progeny size with increasing age is often attributed to a depletion of the female's resources (36, 180, 210, 228). Studies in which maternal diet is manipulated (69, 159) support this hypothesis. Alternatively, decreasing progeny size with increasing maternal age may be adaptive when female clutch size is constrained (11); young females should allocate a larger proportion of their resources to reproduction when their chances of surviving to lay the next clutch are lower. However, this hypothesis assumes that age-specific fecundity does not evolve, an assumption that is unlikely to be realistic.

Maternal age effects on progeny size may reflect a bet-hedging strategy (174); selection favors variation among progeny to ensure that at least some progeny are well suited for future environmental conditions. However, this raises the question of why females do not simply produce the full range of offspring sizes within each clutch or age class. That changes in egg size are sometimes non-adaptive is suggested by the observation that female *Daphnia* produce the size offspring that maximized maternal fitness when they were youngest (24).



In general, eggs laid by older females are less likely to hatch (68, 69, 77, 229) and progeny hatching from these eggs have higher mortality (69, 75, 110, 222; but see 102), produce smaller nymphs/larvae (132), and take longer to reach maturity (69, 77, 137, 222; but see 180) or longer to pupate (102). Sometimes progeny produced by older mothers mature smaller (110, 185) but more often they mature at normal size (69, 77, 102, 180, 222), generally by increasing development time. The sex ratio of progeny may also change as females age (102). These effects on progeny are probably in part mediated by the changes in egg size. However, egg composition (e.g. proportion yolk) also often changes with maternal age (210), such that maternal age effects on progeny cannot be attributed to a decrease in egg size without more careful and creative experimentation.

## CONCLUSIONS

Arthropods exhibit substantial variation in progeny size among species, among individuals within species, and sometimes even among progeny produced by a single female. Many theoretical models have been developed to explain some of this variation, but most start with the same two assumptions as Smith & Fretwell (197)—they assume that progeny fitness increases with increasing progeny size, and that there is a trade-off between progeny size and number. We find that these two assumptions are generally supported by data but that the trade-off between progeny size and number is less apparent when organisms are iteroparous, use adult-acquired resources for reproduction, or provide parental care. This is because variation in total reproductive effort is difficult to quantify for these species. Most models solve for optimal progeny size by assuming that total reproductive effort is constant. However, reproductive effort may vary substantially among individuals and may evolve in response to natural selection. Thus, selection for increased progeny size may lead to increased reproductive effort rather than a decrease in fecundity. This possibility has been examined theoretically but needs to be explored empirically.

Much of the variation in progeny size among species, and among populations within species, appears to have evolved in response to differences in natural selection among environments. Many environmental factors covary with variation in progeny size, and these factors may be the cause of the species or population differences. However, few studies have manipulated progeny environments and demonstrated that the relationship between progeny size and fitness actually differs among environments, and fewer still have demonstrated why selection favors different sized progeny in different environments (e.g. why does selection favor larger eggs at lower temperatures?). Understanding the evolution of intra- and interspecific variation in progeny size will require more empirical studies that identify sources of natural selection within environments and that demonstrate how selection varies among environments.

Much of the variation in progeny size within populations appears to be non-adaptive. For example, smaller females generally lay smaller eggs as an inevitable consequence of phenotypic and genetic correlations between body size and egg size (due to morphological or physiological constraints). However, maternal body size explains a surprisingly small amount of the variation in progeny size within and among populations of many species. Much of the remaining variance is probably also nonadaptive, due to variation in factors such as maternal diet (e.g. food-stressed females generally produce smaller progeny). Yet some species of arthropods, especially crustaceans but also a few insects, exhibit plasticity in progeny size in response to several environmental factors, and much of this plasticity is likely adaptive. Unfortunately, few studies have examined the fitness consequences of plasticity in progeny size, and results of these studies are not always consistent (e.g. comparisons among *Daphnia* studies). The evolution of life history plasticity, including adaptive plasticity in progeny size, is one of the most exciting topics in the study of life histories. We thus suggest that substantially more research effort should be dedicated to understanding the evolution of reaction norms for progeny size.

Variation in size of progeny produced by a single female has been more difficult to explain than variation among females or among populations. Most theoretical models predict that females should produce progeny of a single size. Yet progeny size sometimes varies greatly within families (e.g. changes with female age). We suggest that much of the variation within families is probably nonadaptive. However, some authors have suggested that at least some of the variation within families is an adaptive response to living in a variable environment. At this time, however, there are few experimental studies and too little theoretical work to generalize.

The evolution of progeny size has been extensively modeled by theoretical evolutionary ecologists. However, advances in theory have substantially outpaced empirical data—few empirical studies have progressed much beyond documenting patterns of variation in progeny size within or among population, measuring phenotypic correlations between progeny size and maternal fecundity, or quantifying the relationship between progeny size and a few components of progeny fitness in one environment. We hope that this review will stimulate researchers to examine the specific factors that result in variation in selection on progeny size within and among populations, and how this variation in selection influences the evolution of the patterns that we observe.

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## LITERATURE CITED

1. Albuquerque GS, Tauber MJ, Tauber CA. 1997. Life history adaptations and reproductive costs associated with specialization in predacious insects. *J. Anim. Ecol.* 66:307–17
2. Allan JD. 1984. Life history variation in a freshwater copepod: evidence from population crosses. *Evolution* 38:280–91
3. Anderson J. 1990. The size of spider eggs and estimates of their energy content. *J. Arachnol.* 18:73–78
4. Avelar T. 1993. Egg size in *Drosophila*—standard unit of investment or variable response to environment?—the effect of temperature. *J. Insect Physiol.* 39:283–89
5. Avelar T, Rocha Pit  MT. 1989. Egg size and number in *Drosophila subobscura* under semi-natural conditions. *Evol. Biol.* 3:37–48
6. Azevedo RBR, French V, Partridge L. 1996. Thermal evolution of egg size in *Drosophila melanogaster*. *Evolution* 50:2338–45
7. Azevedo RBR, French V, Partridge L. 1997. Life-history consequences of egg size in *Drosophila melanogaster*. *Am. Nat.* 150:250–82
8. Bakker K. 1969. Selection for rate of growth and its influence on competitive ability of larvae of *Drosophila melanogaster*. *Neth. J. Zool.* 19:541–95
9. Barbosa P, Capinera JL. 1978. Population quality, dispersal and numerical change in the gypsy moth, *Lymantria dispar* (L.). *Oecologia* 36:203–9
10. Beckwith RC. 1982. Effects of constant laboratory temperatures on the Douglas-fir tussock moth (Lepidoptera: Lymantriidae). *Environ. Entomol.* 11:1159–63
11. Begon M, Parker GA. 1986. Should egg size and clutch size decrease with age? *Oikos* 47:293–302
12. Belk D. 1977. Evolution of egg size strategies in fairy shrimps. *Southwest. Nat.* 22:99–105
13. Belk D, Anderson G, Hsu SY. 1990. Additional observations on variations in egg size among populations of *Streptocephalus seali* (Anostraca). *J. Crust. Biol.* 10:128–33
14. Bell G. 1983. Measuring the cost of reproduction III. The correlation structure of the early life history of *Daphnia pulex*. *Oecologia* 60:378–83
15. Berberovic R, Bikar K, Geller W. 1990. Seasonal variability of the embryonic development time of three planktonic crustaceans—dependence on temperature, adult size, and egg weight. *Hydrobiology* 203:127–36
16. Berger A. 1989. Egg weight, batch size and fecundity of the spotted stalk borer, *Chilo partellus* in relation to weight of females and time of oviposition. *Entomol. Exp. Appl.* 50:199–207
17. Bernardo J. 1996. The particular maternal effect of propagule size, especially egg size: patterns, models, quality of evidence and interpretations. *Am. Zool.* 36:216–36
18. Berrigan D. 1991. The allometry of egg size and number in insects. *Oikos* 60:313–21
19. Blackburn TM. 1991. Evidence for a “fast-slow” continuum of life-history traits among parasitoid Hymenoptera. *Funct. Ecol.* 5:65–74
20. Blackenhorn WU, Fairbairn DJ. 1995. Life history adaptation along a latitudinal cline in the water strider *Aquarius remigis* (Heteroptera: Gerridae). *J. Evol. Biol.* 8: 21–41
21. Blau WS. 1981. Life history variation in the black swallowtail butterfly. *Oecologia* 48:116–22

22. Boersma M. 1995. The allocation of resources to reproduction in *Daphnia galeata*: against the odds? *Ecology* 76:1251–61
23. Boersma M. 1997. Offspring size in *Daphnia*: does it pay to be overweight? *Hydrobiologia* 360:79–88
24. Boersma M. 1997. Offspring size and parental fitness in *Daphnia magna*. *Evol. Ecol.* 11:439–50
25. Boersma M, Spaak P, de Meester L. 1998. Predator-mediated plasticity in morphology, life history, and behavior of *Daphnia*: the uncoupling of responses. *Am. Nat.* 152:237–48
26. Boggs CL. 1986. Reproductive strategies of female butterflies: variation in and constraints on fecundity. *Ecol. Entomol.* 11:7–15
27. Braby MF. 1994. The significance of egg size variation in butterflies in relation to host plant quality. *Oikos* 71:119–29
28. Braby MF, Jones RE. 1995. Reproductive patterns and resource allocation in tropical butterflies: influence of adult diet and seasonal phenotype on fecundity, longevity and egg size. *Oikos* 72:189–204
29. Bradshaw WE, Holzapfel CM, O'Neill T. 1993. Egg size and reproductive allocation in the pitcherplant mosquito *Wyeomyia smithii* (Diptera, Culicidae). *J. Med. Entomol.* 30:384–90
30. Brambilla DJ. 1982. Seasonal variation of egg size and number in a *Daphnia pulex* population. *Hydrobiologia* 97:233–48
31. Brittain JE, Lillehammer A, Saltveit SJ. 1984. The effect of temperature on intraspecific variation in egg biology and nymphal size in the stonefly, *Capnia atra* (Plecoptera). *J. Anim. Ecol.* 53:161–69
32. Brody MS, Lawlor LR. 1984. Adaptive variation in offspring size in a terrestrial isopod, *Armadillidium vulgare*. *Oecologia* 61:55–59
33. Burgis MJ. 1967. A quantitative study of reproduction in some species of *Ceriodaphnia* (Crustacea: Cladocera). *J. Anim. Ecol.* 36:61–75
34. Burns CW. 1995. Effects of crowding and different food levels on growth and reproductive investment of *Daphnia*. *Oecologia* 101:234–44
35. Butlin RK, Woodhatch CW, Hewitt GW. 1987. Male spermatophore investment increases female fecundity in a grasshopper. *Evolution* 41:221–25
36. Campbell IM. 1962. Reproductive capacity in the genus *Choristoneura* Led. (Lepidoptera: Tortricidae). I. Quantitative inheritance and genes as controllers of rates. *Can. J. Gen. Cytol.* 4:272–88
37. Carlberg U. 1984. Variation in the egg-size of *Extatosoma tiaratum* (MacLeay) (Insecta: Phasmida). *Zool. Anz.* 212:61–67
38. Carlberg U. 1991. Egg-size variation in *Extatosoma tiaratum* (MacLeay) and its effect on survival and fitness of newly hatched nymphs (Insecta, Phasmida). *Biol. Zent.* 110:163–73
39. Carriere Y, Masaki S, Roff DA. 1997. The coadaptation of female morphology and offspring size: a comparative analysis in crickets. *Oecologia* 110:197–204
40. Carriere Y, Roff DA. 1995. The evolution of offspring size and number: a test of the Smith-Fretwell model in three species of crickets. *Oecologia* 102:389–96
- 40a. Charnov EL, Downhower JF. 1995. A trade-off-invariant life-history rule for optimal offspring size. *Nature* 376:418–19
41. Christiansen FB, Fenchel TM. 1979. Evolution of marine invertebrate reproductive patterns. *Theor. Popul. Biol.* 16:267–82
42. Clarke A. 1979. On living in cold water: K-strategies in Antarctic benthos. *Mar. Biol.* 55:111–19
43. Clarke A, Gore DJ. 1992. Egg size and composition in *Ceratoserolis* (Crustacea, Isopoda) from the Weddell sea. *Polar Biol.* 12:129–34

44. Clarke A, Hopkins CCE, Nilssen EM. 1991. Egg size and reproductive output in the deep-water prawn *Pandalus borealis* Kroyer, 1838. *Funct. Ecol.* 5:724–30
45. Cleuvers M, Goser B, Rattle H-T. 1997. Life-strategy shift by intraspecific interaction in *Daphnia magna*: change in reproduction from quantity to quality. *Oecologia* 110:337–45
46. Clutton-Brock TH. 1991. *The Evolution of Parental Care*. Princeton, NJ: Princeton Univ. Press
- 46a. Corkum LD, Ciborowski JHH, Poulin RG. 1997. Effects of emergence date and maternal size on egg development and sizes of eggs and first-instar nymphs of a semelparous aquatic insect. *Oecologia* 111:69–75
47. Crill WD, Huey RB, Gilchrist GW. 1996. Within- and between-generation effects of temperature on the morphology and physiology of *Drosophila melanogaster*. *Evolution* 50:1205–18
48. Curtsinger JW. 1976. Stabilizing selection in *Drosophila melanogaster*. *J. Hered.* 67:59–60
49. Dangerfield JM. 1997. Growth and survivorship in juvenile woodlice: is birth mass important? *Ecography* 20:132–36
50. Dangerfield JM, Telford SR. 1990. Breeding phenology, variation in reproductive effort and offspring size in a tropical population of the woodlouse *Porcellionides pruinosus*. *Oecologia* 82:251–58
51. Dixon AFG, Kundo R, Kindlmann P. 1993. Reproductive effort and maternal age in iteroparous insects using aphids as a model group. *Funct. Ecol.* 7:267–72
52. Dixon AFG, Wratten SD. 1971. Laboratory studies on aggregation, size and fecundity in the black bean aphid, *Aphis fabae* Scop. *Bull. Entomol. Res.* 61:97–111
53. Dodson GN, Marshall LD. 1984. Male aggression and female egg size in a mate-guarding ambush bug: are they related? *Psyche* 91:193–99
54. Dunlap-Pianka HL. 1979. Ovarian dynamics in *Heliconius* butterflies: correlations among daily oviposition rates, egg weights, and quantitative aspects of oogenesis. *J. Insect Physiol.* 25:741–49
55. Eberhard W. 1979. Rate of egg production by tropical spiders in the field. *Biotropica* 11:292–300
56. Ebert D. 1991. The effect of size at birth, maturation threshold and genetic differences on the life-history of *Daphnia magna*. *Oecologia* 86:243–50
57. Ebert D. 1993. The trade-off between offspring size and number in *Daphnia magna*: The influence of genetic, environmental, and maternal effects. *Arch. Hydrobiol.* 1993:453–73
58. Ebert D. 1994. Fractional resource allocation into few eggs: *Daphnia* as an example. *Ecology* 75:568–71
59. Ebert D, Yampolsky L, Stearns SC. 1993. Genetics of life history in *Daphnia magna*. I. Heritabilities at two food levels. *Heredity* 70:335–43
60. Ebert D, Yampolsky L, van Noordwijk AJ. 1993. Genetics of life history in *Daphnia magna*. II. Phenotypic plasticity. *Heredity* 70:344–52
61. Emler RB, Hoegh-Guldberg O. 1997. Effects of egg size on postlarval performance: experimental evidence from a sea urchin. *Evolution*: 51:141–52
62. Enserink L, Luttmer W, Mass-Diepeveen H. 1990. Reproductive strategy of *Daphnia magna* affects the sensitivity of its progeny in acute toxicity tests. *Aquat. Toxicol.* 17:15–25
63. Ernsting G, Isaaks JA. 1997. Effects of temperature and season on egg size, hatchling size and adult size in *Notiphilus biguttatus*. *Ecol. Entomol.* 22:32–40
64. Fitt GP. 1990. Variation in ovariole number and egg size of species of *Dacus* (Diptera, Tephritidae) and their relation to host specialization. *Ecol. Entomol.* 15:255–64

65. Fitt GP. 1990. Comparative fecundity, clutch size, ovariolo number and egg size of *Dacus tryoni* and *D. jarvisi*, and their relationship to body size. *Entomol. Exp. Appl.* 55:11–21
66. Fitzpatrick SM, Troubridge JT. 1993. Fecundity, number of diapause eggs, and egg size of successive generations of the blackheaded fireworm (Lepidoptera, Tortricidae) on cranberries. *Environ. Entomol.* 22:818–23
67. Forbes LS. 1991. Optimal size and number of offspring in a variable environment. *J. Theor. Biol.* 150:299–304
- 67a. Forbes LS. 1999. Within-clutch variation in propagule size: the double-default model. *Oikos* 85:146–50
68. Forrest TG. 1986. Oviposition and maternal investment in mole crickets (Orthoptera: Gryllotalpidae): effects of season, size, and senescence. *Ann. Entomol. Soc. Am.* 79:918–24
69. Fox CW. 1993. The influence of maternal age and mating frequency on egg size and offspring performance in *Callosobruchus maculatus* (Coleoptera: Bruchidae). *Oecologia* 96:139–46
70. Fox CW. 1994. The influence of egg size on offspring performance in the seed beetle, *Callosobruchus maculatus*. *Oikos* 71:321–25
71. Fox CW. 1994. Maternal and genetic influences on egg size and larval performance in a seed beetle: multigenerational transmission of a maternal effect? *Heredity* 73:509–17
72. Fox CW. 1997. Egg size manipulations in the seed beetle, *Stator limbatus*: consequences for progeny growth. *Can. J. Zool.* 75:1465–73
73. Fox CW. 1997. The ecology of body size in a seed beetle, *Stator limbatus*: persistence of environmental variation across generations? *Evolution* 51:1005–10
74. Fox CW. 2000. Natural selection on seed beetle egg size in the field and the lab: variation among environments. *Ecology*. In press
75. Fox CW, Czesak ME, Mousseau TA, Roff DA. 1999. The evolutionary genetics of an adaptive maternal effect: egg size plasticity in a seed beetle. *Evolution*. In press
76. Fox CW, Czesak ME, Savalli UM. 1999. Environmentally-based maternal effects on development time in the seed beetle, *Stator pruininus* (Coleoptera: Bruchidae): consequences of larval density. *Environ. Entomol.* 28:217–23
77. Fox CW, Dingle H. 1994. Dietary mediation of maternal age effects on offspring performance in a seed beetle (Coleoptera: Bruchidae). *Funct. Ecol.* 8:600–6
78. Fox CW, Mousseau TA. 1996. Larval host plant affects the fitness consequences of egg size in the seed beetle *Stator limbatus*. *Oecologia* 107:541–48
79. Fox CW, Savalli UM. 1998. Inheritance of environmental variation in body size: Superparasitism of seeds affects progeny and grandprogeny body size via a non-genetic maternal effect. *Evolution* 52:172–82
80. Fox CW, Savalli UM. 1999. Maternal effects mediate diet expansion in a seed-feeding beetle. *Ecology*. In press
81. Fox CW, Thakar MS, Mousseau TA. 1997. Egg size plasticity in a seed beetle: an adaptive maternal effect. *Am. Nat.* 149:149–63
82. Fox CW, Waddell KJ, des Lauriers J, Mousseau TA. 1997. Seed beetle survivorship, growth and egg size plasticity in a paloverde hybrid zone. *Ecol. Entomol.* 22:416–24
83. García-Barros E. 1992. Evidence for geographic variation of egg size and fecundity in a satyrine butterfly, *Hipparchia semele* (L.) (Lepidoptera, Nymphalidae-Satyrinae). *Graellsia* 48:45–52
84. García-Barros E. 1994. Egg size variation in European satyrine butterflies (Nymphalidae, Satyrinae). *Biol. J. Linn. Soc.* 51:309–24
85. García-Barros E, Munguira ML. 1997. Uncertain branch lengths, taxonomic sampling error, and the egg to body size allometry in temperate butterflies (Lepidoptera). *Biol. J. Linn. Soc.* 61:201–21
86. Gilbert F. 1990. Size, phylogeny and life

- history evolution of feeding specialization in insect predators. In *Insect Life Cycles, Genetics, Evolution and Coordination*, ed. F Gilbert, pp. 101–24. Berlin: Springer-Verlag
87. Glazier DS. 1992. Effects of food, genotype, and maternal size and age on offspring investment in *Daphnia magna*. *Ecology* 73:910–26
88. Glazier DS. 1998. Does body storage act as a food-availability cue for adaptive adjustment of egg size and number in *Daphnia magna*? *Freshw. Biol.* 40:87–92
89. Gliwicz ZM, Guisande C. 1992. Family planning in *Daphnia*: resistance to starvation in offspring born to mothers at different food levels. *Oecologia* 91:463–67
90. Green J. 1956. Growth, size and reproduction in *Daphnia* (Crustacea: Cladocera). *Proc. Zool. Soc. London* 126:173–204
91. Green J. 1966. Seasonal variation in egg production by Cladocera. *J. Anim. Ecol.* 35:77–104
92. Guisande C, Gliwicz ZM. 1992. Egg size and clutch size in two *Daphnia* species grown at different food levels. *J. Plankton Res.* 14:997–1007
93. Guisande C, Sanchez J, Maneiro I, Miranda A. 1996. Trade-off between offspring number and offspring size in the marine copepod *Euterpina acutifrons* at different food concentrations. *Mar. Ecol. Prog. Ser.* 143:37–44
94. Guntrip J, Sibly RM, Smith RH. 1997. Controlling resource acquisition to reveal a life history trade-off: egg mass and clutch size in an iteroparous seed predator, *Prostephanus truncatus*. *Ecol. Entomol.* 22:264–70
95. Gwynne DT. 1984. Courtship feeding increases female reproductive success in bushcrickets. *Nature* 307:361–63
96. Gwynne DT. 1988. Courtship feeding and the fitness of female katydids (Orthoptera: Tettigoniidae). *Evolution* 42:545–55
97. Hanazato T, Dodson SI. 1995. Synergistic effects of low oxygen concentration, predator kairomone, and a pesticide on the cladoceran *Daphnia pulex*. *Limnol. Oceanogr.* 40:571–77
98. Hancock MA. 1998. The relationship between egg size and embryonic and larval development in the freshwater shrimp *Paratya australiensis* Kemp (Decapoda: Atyidae). *Freshw. Biol.* 39:715–23
99. Hard JJ, Bradshaw WE. 1993. Reproductive allocation in the western tree-hole mosquito, *Aedes sierrensis*. *Oikos* 66:55–65
100. Hart MW. 1995. What are the costs of small egg size for a marine invertebrate with feeding planktonic larvae? *Am. Nat.* 146:415–26
101. Hart RC, McLaren IA. 1978. Temperature acclimation and other influences on embryonic duration in the copepod, *Pseudocalanus* sp. *Mar. Biol.* 45:23–30
102. Harvey GT. 1977. Mean weight and rearing performance of successive egg clusters of eastern spruce budworm (Lepidoptera: Tortricidae). *Can. Entomol.* 109:487–96
103. Harvey GT. 1983. A geographical cline in egg weights in *Choristoneura fumiferana* (Lepidoptera: Tortricidae) and its significance in population dynamics. *Can. Entomol.* 115:1103–8
104. Harvey GT. 1983. Environmental and genetic effects on mean egg weight in spruce budworm (Lepidoptera: Tortricidae). *Can. Entomol.* 115:1109–17
105. Harvey GT. 1985. Egg weight as a factor in the overwintering survival of spruce budworm (Lepidoptera: Tortricidae) larvae. *Can. Entomol.* 117:1451–61
106. Hill CJ. 1989. The effect of adult diet on the biology of butterflies 2. The common crow butterfly, *Euploea core corinna*. *Oecologia* 81:258–66
107. Hill CJ, Pierce NE. 1989. The effect of adult diet on the biology of butterflies. I. The common imperial blue, *Jalmenus evagoras*. *Oecologia* 81: 249–57
108. Holloway GJ, Smith RH, Wrelton AE,

- King PE, Li LL, Menendez GT. 1987. Egg size and reproductive strategies in insects infesting stored-products. *Funct. Ecol.* 1:229–35
109. Honěk A. 1987. Regulation of body size in a heteropteran bug, *Pyrrhocoris apterus*. *Entomol. Exp. Appl.* 44:257–62
110. Honěk A. 1992. Female size, reproduction and progeny size in *Pyrrhocoris apterus* (Heteroptera, Pyrrhocoridae). *Acta Entomol. Bohemoslov* 89:169–78
111. Huey RB. 1995. Within- and between-generation effects of temperature on early fecundity of *Drosophila melanogaster*. *Heredity* 74:216–23
112. Johnson LK. 1982. Sexual selection in brentid weevils. *Evolution* 36:251–62
113. Jones RE, Hart JR, Bull GD. 1982. Temperature, size and egg production in the cabbage butterfly, *Pieris rapae* L. *Aust. J. Zool.* 30:223–32
114. Juliano SA. 1985. The effects of body size on mating and reproduction in *Brachinus lateralis* (Coleoptera: Carabidae). *Ecol. Entomol.* 10:271–80
115. Kaplan RH, Cooper WS. 1988. On the evolution of coin-flipping plasticity: a response to McGinley, Temme, and Geber. *Am. Nat.* 132:753–55
116. Karlsson B. 1987. Variation in egg weight, oviposition rate and reproductive reserves with female age in a natural population of the speckled wood butterfly, *Pararge aegeria*. *Ecol. Entomol.* 12:473–76
117. Karlsson B, Wickman P-O. 1990. Increase in reproductive effort as explained by body size and resource allocation in the speckled wood butterfly, *Pararge aegeria* (L.). *Funct. Ecol.* 4:609–17
118. Karlsson B, Wiklund C. 1984. Egg weight variation and lack of correlation between egg weight and offspring fitness in the wall brown butterfly *Lasiommata megera*. *Oikos* 43:376–85
119. Karlsson B, Wiklund C. 1985. Egg weight variation in relation to egg mortality and starvation endurance of newly hatched larvae in some satyrid butterflies. *Ecol. Entomol.* 10:205–11
120. Kasule FK. 1991. Egg size increases with maternal age in the cotton stainer bugs *Dysdercus fasciatus* and *D. cardinalis* (Hemiptera: Pyrrhocoridae). *Ecol. Entomol.* 16:345–49
121. Kawamura N. 1990. Is the egg size determining gene, ESD, on the W-chromosome identical with the sex-linked giant egg gene, GE, in the silkworm? *Genetica* 81:205–10
122. Kawecki TJ. 1995. Adaptive plasticity of egg size in response to competition in the cowpea weevil, *Callosobruchus maculatus* (Coleoptera: Bruchidae). *Oecologia* 102: 81–85
123. Kerfoot WC. 1974. Egg size cycle of a cladoceran. *Ecology* 55:1259–70
124. Kessler A. 1971. Relation between egg production and food consumption in species of the genus *Pardosa* (Lycosidae, Araneae) under experimental conditions of food abundance and shortage. *Oecologia* 8:93–109
125. Kimura K, Masaki S. 1977. Brachypterism and seasonal adaptation in *Orgyia thyellina* Butler (Lepidoptera: Lymantriidae). *Kontyû* 45:97–106
126. Kimura K, Tsubaki Y. 1985. Egg weight variation associated with female age in *Pieris rapae crucivora* Boisduval (Lepidoptera: Pieridae). *Appl. Entomol. Zool.* 20:500–1
127. Kolding S, Fenchel TM. 1981. Patterns of reproduction in different populations of five species of the amphipod genus *Gammarus*. *Oikos* 37:167–72
128. Kozłowski J. 1996. Optimal initial size and adult size of animals: consequences for macroevolution and community structure. *Am. Nat.* 147:101–14
129. Lamb RJ, Smith SM. 1980. Comparisons of egg size and related life-history characteristics for two predaceous tree-hole mosquitos (*Toxorhynchites*). *Can. J. Zool.* 58:2065–70



130. Lampert W. 1993. Phenotypic plasticity of the size at first reproduction in *Daphnia*: the importance of maternal size. *Ecology* 74:1455–66
131. Landa K. 1992. Adaptive seasonal variation in grasshopper offspring size. *Evolution* 46:1553–58
132. Larsson FK. 1989. Female longevity and body size as predictors of fecundity and egg length in *Graphosoma lineatum* L. *Dtsch. Entomol. Z.* 36:329–34
133. Larsson FK. 1990. Female body size relationships with fecundity and egg size in two solitary species of fossorial Hymenoptera (Colletidae and Sphecidae). *Entomol. Gen.* 15:167–71
134. Larsson FK, Kustvall V. 1990. Temperature reverses size-dependent male mating success of a cerambycid beetle. *Funct. Ecol.* 4:85–90
135. Lawlor LR. 1976. Parental investment and offspring fitness in the terrestrial isopod *Armadillidium vulgare* (Latr.), (Crustacea: Oniscoidea). *Evolution* 30:775–85
136. Leather SR, Burnand AC. 1987. Factors affecting life-history parameters of the pine beauty moth, *Panolis flammea* (D&S): the hidden costs of reproduction. *Funct. Ecol.* 1:331–38
137. Leonard DE. 1970. Intrinsic factors causing qualitative changes in populations of *Porthetria dispar* (Lepidoptera: Lymantriidae). *Can. Entomol.* 102:239–49
138. LePrince DJ, Foil LD. 1993. Relationships among body size, blood meal size, egg volume, and egg production of *Tabanus fuscicostatus* (Diptera: Tabanidae). *J. Med. Entomol.* 30:865–71
139. Lynch M. 1980. The evolution of cladoceran life histories. *Q. Rev. Biol.* 55:23–42
140. Lynch M. 1984. The limits to life history evolution in *Daphnia*. *Evolution* 38:465–82
141. Lynch M. 1989. The life history consequences of resource depression in *Daphnia pulex*. *Ecology* 70:246–56
142. Lynch M, Pfrender M, Spitze K, Lehman N, Hicks J, et al. 1999. The quantitative and molecular genetic architecture of a subdivided species. *Evolution* 53:100–10
143. Maeta Y, Takahashi K, Shimada N. 1998. Host body size as a factor determining the egg complement of Strepsiptera, an insect parasite. *J. Insect Morphol. Embryol.* 27:27–37
144. Magalhães C, Walker I. 1988. Larval development and ecological distribution of Central Amazonian palaemonid shrimp (Decapoda, Caridea). *Crustaceana* 55:279–92
145. Mappes J, Mappes T, Lappalainen T. 1997. Unequal maternal investment in offspring quality in relation to predation risk. *Evol. Ecol.* 11:237–43
146. Marshall LD. 1990. Intraspecific variation in reproductive effort by female *Parapediasia teterella* (Lepidoptera: Pyralidae) and its relation to body size. *Can. J. Zool.* 68:44–48
147. Marshall SD, Gittleman JL. 1994. Clutch size in spiders: is more better? *Funct. Ecol.* 8:118–24
148. Mashiko K. 1982. Differences in both the egg size and the clutch size of the freshwater prawn *Palaemon paucidens* de Haan in the Sagami river. *Jpn. J. Ecol.* 32:445–51
149. Mashiko K. 1983. Differences in egg and clutch sizes of the prawn *Macrobrachium nipponense* (de Haan) between brackish and freshwaters of a river. *Zool. Mag.* 92:1–9
150. Mashiko K. 1990. Diversified egg and clutch sizes among local populations of the fresh-water prawn *Macrobrachium nipponense* (de Haan). *J. Crust. Biol.* 10:306–14
151. Mashiko K. 1992. Genetic egg and clutch size variations in freshwater prawn populations. *Oikos* 63:454–58
152. McGinley MA. 1989. The influence of a positive correlation between clutch size and offspring fitness on the optimal offspring size. *Evol. Ecol.* 3:150–56

153. McKee D, Ebert D. 1996. The interactive effects of temperature, food level and maternal phenotype on offspring size in *Daphnia magna*. *Oecologia* 107:189–96
154. McLain DK, Mallard SD. 1991. Sources and adaptive consequences of egg size variation in *Nezara viridula* (Hemiptera: Pentatomidae). *Psyche* 98:135–64
155. McLaren IA. 1965. Some relationships between temperature and egg size, body size, development rate, and fecundity, of the copepod *Pseudocalanus*. *Limnol. Oceanogr.* 10:528–38
- 155a. Mohaghegh J, De Clerco P, Tirry L. 1998. Effects of maternal age and egg weight on development time and body weight of offspring of *Podisus maculiventris* (Heteroptera: Pentatomidae). *Ann. Entomol. Am.* 91:315–322.
156. Moore M, Folt C. 1993. Zooplankton body size and community structure: effects of thermal and toxicant stress. *TREE* 8:178–83
157. Moore RA, Singer MC. 1987. Effects of maternal age and adult diet on egg weight in the butterfly *Euphydryas editha*. *Ecol. Entomol.* 12:401–8
158. Mura G. 1991. Additional remarks on cyst morphometrics in anostracans and its significance. 1. Egg size. *Crustaceana* 61:241–52
159. Murphy DD, Launer AE, Ehrlich PR. 1983. The role of adult feeding in egg production and population dynamics of the checkerspot butterfly *Euphydryas editha*. *Oecologia* 56:257–63
160. Nakasuji F. 1987. Egg size of skippers (Lepidoptera: Hesperidae) in relation to their host specificity and to leaf toughness of host plants. *Ecol. Res.* 2:175–83
161. Nakasuji F, Kimura M. 1984. Seasonal polymorphism of egg size in a migrant skipper, *Parnara guttata guttata* (Lepidoptera, Hesperidae). *Kontyû* 52:253–59
162. Newton C, Dixon AFG. 1990. Embryonic growth and birth rate of the offspring of apterous and alate aphids: a cost of dispersal. *Entomol. Exp. Appl.* 55:223–29
163. Nishino M. 1990. Geographic variation of body size, brood size and egg size of a freshwater shrimp, *Palaemon paucidens* de Haan with some discussion on brood habitat. *Jpn. J. Limnol.* 41:185–202
164. Oberhauser KS. 1997. Fecundity, life-span and egg mass in butterflies: Effects of male-derived nutrients and female size. *Funct. Ecol.* 11:166–75
165. Odinetz-Collart O, Rabelo H. 1996. Variation in egg size of the fresh-water prawn *Macrobrachium amazonicum* (Decapoda, Palaemonidae). *J. Crust. Biol.* 16:684–88
166. O'Neill KM, Skinner SW. 1990. Ovarian egg size and number in relation to female size in five species of parasitoid wasps. *J. Zool.* 220:115–22
167. Pagel M. 1999. Mother and father in surprise genetic agreement. *Nature* 397:19–20
168. Palmer JO. 1985. Life-history consequences of body-size variation in the milkweed leaf beetle, *Labidomera clivicollis* (Coleoptera: Chrysomelidae). *Ann. Entomol. Soc. Am.* 78:603–8
169. Parker GA, Begon M. 1986. Optimal egg size and clutch size: effects of environment and maternal phenotype. *Am. Nat.* 128:573–92
170. Parsons PA. 1964. Egg lengths in *Drosophila melanogaster* and correlated responses to selection. *Genetica* 35:175–81
171. Parsons PA. 1964. Parental age and the offspring. *Q. Rev. Biol.* 39:258–75
172. Perrin N. 1988. Why are offspring born larger when it is colder? Phenotypic plasticity for offspring size in the cladoceran *Simocephalus vetulus* (Müller). *Funct. Ecol.* 2:283–88
173. Perrin N. 1989. Population density and offspring size in the cladoceran *Simocephalus vetulus* (Müller.). *Funct. Ecol.* 3:29–36
174. Philippi T, Seger J. 1989. Hedging one's evolutionary bets, revisited. *TREE* 4:41–44

175. Poulin R. 1995. Clutch size and egg size in free-living and parasitic copepods: a comparative analysis. *Evolution* 49:325–36
176. Poulin R, Hamilton WJ. 1997. Ecological correlates of body size and egg size in parasitic *Ascothoracida* and *Rhizocephala* (Crustacea). *Acta Oecol.* 18:621–35
177. Price PW. 1973. Reproductive strategies in parasitoid wasps. *Am. Nat.* 107:684–93
178. Reavey D. 1992. Egg size, first instar behaviour and the ecology of Lepidoptera. *J. Zool. London* 227:277–97
179. Reede T. 1997. Effects of neonate size and food concentration on the life history responses of a clone of the hybrid *Daphnia hyalina* X *galeata* to fish kairomones. *Freshw. Biol.* 37:389–96
180. Richards LJ, Myers JH. 1980. Maternal influences on size and emergence time of the cinnabar moth. *Can. J. Zool.* 58:1452–57
181. Robertson AL. 1988. Life histories of some species of Chydoridae (Cladocera: Crustacea). *Freshw. Biol.* 20:75–84
182. Roff DA. 1992. *The Evolution of Life Histories: Theory and Analysis*. New York: Chapman & Hall
183. Rosenheim JA. 1996. An evolutionary argument for egg limitation. *Evolution* 50:2089–94
184. Rossiter MC. 1991. Maternal effects generate variation in life history: consequences of egg weight plasticity in the gypsy moth. *Funct. Ecol.* 5:386–93
185. Ruohomäki K, Hanhimäki S, Haukioja E. 1993. Effects of egg size, laying order and larval density on performance of *Epirrita autumnata* (Lep, Geometridae). *Oikos* 68:61–66
186. Savalli UM, Fox CW. 1998. Sexual selection and the fitness consequences of male body size in the seed beetle *Stator limbatus*. *Anim. Behav.* 55:473–83
- 186a. Schwarzkopf L, Blows MW, Caley MJ. 1999. Life history consequences of divergent selection on egg size in *Drosophila melanogaster*. *Am. Nat.* 29:333–40
187. Sheader M. 1996. Factors influencing egg size in the gammarid amphipod *Gammarus insensibilis*. *Mar. Biol.* 124:519–26
188. Shine R. 1989. Alternative models for the evolution of offspring size. *Am. Nat.* 134:311–17
189. Sibly R, Calow P. 1986. *Physiological Ecology of Animals: An Evolutionary Approach*. Oxford: Blackwell Sci.
190. Sibly R, Monk K. 1987. A theory of grasshopper life cycles. *Oikos* 48:186–94
191. Deleted in proof
192. Simmons LW. 1990. Nuptial feedings in tettigoniids: male costs and the rates of fecundity increase. *Behav. Ecol. Sociobiol.* 27:43–47
193. Simpson MR. 1995. Covariation of spider egg and clutch size—the influence of foraging and parental care. *Ecology* 76:795–800
194. Sinervo B. 1993. The effect of offspring size on physiology and life history. *BioScience* 43:210–18
195. Sinervo B, McEdward LR. 1988. Developmental consequences of an evolutionary change in egg size: an experimental test. *Evolution* 42:885–89
196. Skadsheim A. 1984. Coexistence and reproductive adaptations of amphipods: the role of environmental heterogeneity. *Oikos* 43:94–103
197. Smith CC, Fretwell SD. 1974. The optimal balance between size and number of offspring. *Am. Nat.* 108:499–506
198. Solbreck C. 1986. Wing and flight muscle polymorphism in a lygaeid bug, *Horvathiolus gibbicollis*: determinants and life history consequences. *Ecol. Entomol.* 11:435–44
199. Solbreck C, Olsson R, Anderson DB, Forare J. 1989. Size, life history and responses to food shortage in two geographical strains of a seed bug *Lygaeus equestris*. *Oikos* 55:387–96

200. Sota T. 1993. Response to selection for desiccation resistance in *Aedes albopictus* eggs (Diptera: Culicidae). *Appl. Entomol. Zool.* 28:161–68
201. Sota T, Mogi M. 1992. Interspecific variation in desiccation survival time of *Aedes (Stegomyia)* mosquito eggs is correlated with habitat and egg size. *Oecologia* 90:353–58
202. Steinwascher K. 1984. Egg size variation in *Aedes aegypti*: relationship to body size and other variables. *Am. Midl. Nat.* 112:76–84
203. Deleted in proof
204. Stewart LA, Hemptinne J-L, Dixon AFG. 1991. Reproductive tactics of ladybird beetle: relationships between egg size, ovariole number and developmental time. *Funct. Ecol.* 5:380–85
205. Strong DR. 1972. Life history variation among populations of an amphipod (*Hyalella azteca*). *Ecology* 53:1103–11
206. Styan CA. 1998. Polyspermy, egg size, and the fertilization kinetics of free-spawning marine invertebrates. *Am. Nat.* 152:290–97
207. Svard L, Wiklund C. 1988. Fecundity, egg weight and longevity in relation to multiple matings in females of the monarch butterfly. *Behav. Ecol. Sociobiol.* 23:39–43
208. Tanaka S, Wolda H. 1987. Seasonal wing length dimorphism in a tropical seed bug: ecological significance of the short-winged form. *Oecologia* 75:559–65
209. Tauber CA, Tauber MJ, Tauber MJ. 1991. Egg size and taxon: their influence on survival and development of chrysopid hatchlings after food and water deprivation. *Can. J. Zool.* 69:2644–50
210. Telfer WH, Rutberg LD. 1960. Effects of blood protein depletion on the growth of the oocytes in the cecropia moth. *Biol. Bull.* 118:352–66
211. Telford SR, Dangerfield JM. 1995. Offspring size variation in some southern African woodlice. *Afr. J. Ecol.* 33:236–41
212. Tessier AJ, Consolatti NL. 1989. Variation in offspring size in *Daphnia* and consequences for individual fitness. *Oikos* 56:269–76
213. Tessier AJ, Consolatti NL. 1991. Resource quality and offspring quality in *Daphnia*. *Ecology* 72:468–78
214. Toda S, Fujisaki K, Nakasuji F. 1995. The influence of egg size on development of the bean bug, *Riptortus clavatus* Thunberg (Heteroptera: Coreidae). *Appl. Entomol. Zool.* 30:485–87
215. Trubetskova I, Lampert W. 1995. Egg size and egg mass of *Daphnia magna*—response to food availability. *Hydrobiologia* 307:139–45
216. Vahed K. 1998. The function of nuptial feeding in insects: a review of empirical studies. *Biol. Rev.* 73:43–78
217. Villet M. 1990. Qualitative relations of egg size, egg production and colony size in some ponerine ants (Hymenoptera, Formicidae). *J. Nat. Hist.* 24:1321–31
218. Wall R. 1990. Ovarian aging in tsetse flies (Diptera: Glossinidae)—interspecific differences. *Bull. Entomol.* 12:109–14
219. Wallin H, Chiverton PA, Ekblom BS, Borg A. 1992. Diet, fecundity and egg size in some polyphagous predatory carabid beetles. *Entomol. Exp. Appl.* 65:129–40
220. Walsh CJ. 1993. Larval development of *Paratya australiensis* Kemp, 1917 (Decapoda: Caridea: Atyidae), reared in the laboratory, with comparisons of fecundity and egg and larval size between estuarine and riverine environments. *J. Crust. Biol.* 13:456–80
221. Warren DC. 1924. Inheritance of egg size in *Drosophila melanogaster*. *Genetics* 9:41–69
222. Wasserman SS, Asami T. 1985. The effect of maternal age upon fitness of progeny in the southern cowpea weevil, *Callosobruchus maculatus*. *Oikos* 45:191–96
223. Wasserman SS, Mitter C. 1978. The rela-

- tionship of body size to breadth of diet in some Lepidoptera. *Ecol. Entomol.* 3:155–60
224. Webber LG. 1955. The relationship between larval and adult size of the Australian sheep blowfly, *Lucilia cuprina*. *Aust. J. Zool.* 3:346–53
225. Weigensberg I, Carriere Y, Roff DA. 1998. Effects of male genetic contribution and paternal investment to egg and hatchling size in the cricket, *Gryllus firmus*. *J. Evol. Biol.* 11:135–46
226. Wickman P-O, Karlsson B. 1987. Changes in egg color, egg weight and oviposition rate with the number of eggs laid by wild females of the small heath butterfly, *Coenonympha pamphilus*. *Ecol. Entomol.* 12:109–14
227. Wiklund C, Karlsson B. 1984. Egg size variation in satyrid butterflies: adaptive vs. historical, “Bauplan,” and mechanistic explanations. *Oikos* 43:391–400
228. Wiklund C, Karlsson B, Forsberg J. 1987. Adaptive versus constraint explanations for egg-to-body size relationships in two butterfly families. *Am. Nat.* 130:828–38
229. Wiklund C, Persson A. 1983. Fecundity, and the relation of egg weight to offspring fitness in the speckled wood butterfly *Pararge aegeria*, or why don't butterfly females lay more eggs? *Oikos* 40:53–63
230. Willows RI. 1987. Intrapopulation variation in the reproductive characteristics of two populations of *Ligia oceanica* (Crustacea: Oniscidea). *J. Anim. Ecol.* 56:331–40
- 230a. Winkler DW, Wallin K. 1987. Offspring size and number: a life history model linking effort per offspring and total effort. *Am. Nat.* 129:708–20
231. Wittmann KJ. 1981. On the breeding biology and physiology of marsupial development in Mediterranean *Leptomysis* (Mysidacea: Crustacea) with special reference to the effects of temperature and egg size. *J. Exp. Mar. Biol. Ecol.* 53:261–79
232. Yamplosky LY, Scheiner SM. 1996. Why larger offspring at lower temperatures? A demographic approach. *Am. Nat.* 147:86–100

